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Draft Genome Sequence of a Urinary Isolate of *Lactobacillus crispatus*

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While *Lactobacillus crispatus* contributes to the stability of normal vaginal microbiota, its role in urinary health remains unclear. As part of an on-going attempt to characterize the female urinary microbiota, we report the genome sequence of an *L. crispatus* strain isolated from a woman displaying no lower urinary tract symptoms.

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As part of an attempt to characterize the newly discovered female urinary microbiota (1–8), we report the genome sequence and annotation of a strain of *Lactobacillus crispatus* isolated from the bladder of an adult female. This is the first genome report of a urinary isolate of *L. crispatus*, a species associated with bladder health (8).

Using the expanded spectrum version (9) of the enhanced quantitative urine culture protocol (2), *L. crispatus* strain C037 was isolated from a healthy female not displaying any urinary symptoms. The strain was subcultured to purity, analyzed by matrix-assisted laser desorption/ionization-time-of-flight mass spectrometry, and pure cultures were stored at -80°C in 2 mL CryoSaver Brucella broth with 10% glycerol, no beads, cryovials (Hardy Diagnostics). For genome extraction, the preserved pure culture isolate was grown on 5% sheep blood agar (BDBBL™ prepared plated media) under 5% CO_2 at 35°C for 24 h.

To extract genomic DNA, cells were resuspended in 0.5 mL DNA extraction buffer (20 mM tris-Cl, 2 mM EDTA, 1.2% Triton X-100, pH 8) followed by addition of 50 μL lysozyme (20 mg/mL), 30 μL mutanolysin, and 5 μL RNase (10 mg/mL). After a 1-h incubation at 37°C , 80 μL 10% SDS and 20 μL proteinase K were added followed by a 2-h incubation at 55°C . Then, 210 μL of 6M NaCl and 700 μL phenol-chloroform were added. After a 30-min incubation with rotation, the solution was centrifuged at 13,500 rpm for 10 min, and the aqueous phase extracted. An equivalent volume of isopropanol was added; after a 10-min incubation, the solution was centrifuged at 13,500 rpm for 10 min. The supernatant was decanted and the DNA pellet precipitated using 600 μL 70% ethanol. Following ethanol evaporation, the DNA pellet was resuspended in tris-EDTA and stored at -20°C .

Genomic DNA was diluted in water to a concentration of 0.2 ng/ μL . Library preparation was performed using the Nextera XT DNA library preparation kit (Illumina) according to manufacturer's instructions with 1 ng of input DNA. The isolate was sequenced twice, on two separate runs, using the Illumina MiSeq platform and the MiSeq reagent kit v2 (300-cycles). Sequence assembly was performed using Velvet (10) ($k = 99$) followed by SSPACE (11) for scaffolding. *L. crispatus* C037 was assembled into

96 scaffolds with a genome coverage of $113\times$. The scaffolds include 2.147 Mbp of sequence with a G+C content of 36.6%. Gene annotations were performed using GLIMMER (12) and tRNA-Scan (13) identifying 2,096 protein coding genes, 65 RNA (tRNA and rRNA) genes, and four clustered regularly interspaced short palindromic repeats (CRISPR) (14). The 16S rRNA gene sequence of the urinary isolate C037 was identical to that of the species' type strain *L. crispatus* ST1 (NR_074986), an avian enteric strain. One scaffold (10,704 bp in length) produced a hit to the 16,663 bp *L. crispatus* plasmid pLc17 (KR052811); while a significant proportion of the plasmid sequence was detected, a complete, circularized assembly was not possible.

Accession number(s). The draft whole-genome project for *L. crispatus* C037 has been deposited at DDBJ/EMBL/GenBank under accession number **MAKH00000000**.

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REFERENCES

- Fouts DE, Pieper R, Szpakowski S, Pohl H, Knoblach S, Suh MJ, Huang ST, Ljungberg I, Sprague BM, Lucas SK, Torralba M, Nelson KE, Groah SL. 2012. Integrated next-generation sequencing of 16S rDNA and metaproteomics differentiate the healthy urine microbiome from asymptomatic bacteriuria in neuropathic bladder associated with spinal cord injury. *J Transl Med* 10:174. <http://dx.doi.org/10.1186/1479-5876-10-174>.
- Hilt EE, McKinley K, Pearce MM, Rosenfeld AB, Zilliox MJ, Mueller ER, Brubaker L, Gai X, Wolfe AJ, Schreckenberger PC. 2014. Urine is not sterile: use of enhanced urine culture techniques to detect resident bacterial flora in the adult female bladder. *J Clin Microbiol* 52:871–876. <http://dx.doi.org/10.1128/JCM.02876-13>.
- Khasriya R, Sathiananthamoorthy S, Ismail S, Kelsey M, Wilson M,

- Rohn JL, Malone-Lee J. 2013. Spectrum of bacterial colonization associated with urothelial cells from patients with chronic lower urinary tract symptoms. *J Clin Microbiol* 51:2054–2062. <http://dx.doi.org/10.1128/JCM.03314-12>.
4. Lewis DA, Brown R, Williams J, White P, Jacobson SK, Marchesi JR, Drake MJ. 2013. The human urinary microbiome; bacterial DNA in voided urine of asymptomatic adults. *Front Cell Infect Microbiol* 3:41. <http://dx.doi.org/10.3389/fcimb.2013.00041>.
 5. Nienhouse V, Gao X, Dong Q, Nelson DE, Toh E, McKinley K, Schreckenberger P, Shibata N, Fok CS, Mueller ER, Brubaker L, Wolfe AJ, Radek KA. 2014. Interplay between bladder microbiota and urinary antimicrobial peptides: mechanisms for human urinary tract infection risk and symptom severity. *PLoS One* 9:e114185. <http://dx.doi.org/10.1371/journal.pone.0114185>.
 6. Siddiqui H, Nederbragt AJ, Lagesen K, Jeansson SL, Jakobsen KS. 2011. Assessing diversity of the female urine microbiota by high throughput sequencing of 16S rDNA amplicons. *BMC Microbiol* 11:244. <http://dx.doi.org/10.1186/1471-2180-11-244>.
 7. Wolfe AJ, Toh E, Shibata N, Rong R, Kenton K, Fitzgerald M, Mueller ER, Schreckenberger P, Dong Q, Nelson DE, Brubaker L. 2012. Evidence of uncultivated bacteria in the adult female bladder. *J Clin Microbiol* 50:1376–1383. <http://dx.doi.org/10.1128/JCM.05852-11>.
 8. Pearce MM, Hilt EE, Rosenfeld AB, Zilliox MJ, Thomas-White K, Fok C, Kliethermes S, Schreckenberger PC, Brubaker L, Gai X, Wolfe AJ. 2014. The female urinary microbiome: a comparison of women with and without urgency urinary incontinence. *mBio* 5:e01283-14. <http://dx.doi.org/10.1128/mBio.01283-14>.
 9. Price TK, Dune T, Hilt EE, Thomas-White KJ, Kliethermes S, Brincat C, Brubaker L, Wolfe AJ, Mueller ER, Schreckenberger PC. 2016. The clinical urine culture: enhanced techniques improve detection of clinically relevant microorganisms. *J Clin Microbiol* 54:1216–1222. <http://dx.doi.org/10.1128/JCM.00044-16>.
 10. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
 11. Boetzer M, Henkel CV, Jansen HJ, Butler D, Pirovano W. 2011. Scaffolding pre-assembled contigs using SSPACE. *Bioinformatics* 27:578–579. <http://dx.doi.org/10.1093/bioinformatics/btq683>.
 12. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with glimmer. *Bioinformatics* 23:673–679. <http://dx.doi.org/10.1093/bioinformatics/btm009>.
 13. Schattner P, Brooks AN, Lowe TM. 2005. The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs. *Nucleic Acids Res* 33:W686–W689. <http://dx.doi.org/10.1093/nar/gki366>.
 14. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a web tool to identify clustered regularly interspaced short palindromic repeats. *Nucleic Acids Res* 35:W52–W57. <http://dx.doi.org/10.1093/nar/gkm360>.