

Loyola University Chicago [Loyola eCommons](https://ecommons.luc.edu/) 

[Bioinformatics Faculty Publications](https://ecommons.luc.edu/bioinformatics_facpub) Faculty Publications and Other Works by **Department** 

8-12-2020

# Comparative Genomic Study of Lactobacillus jensenii and the Newly Defined Lactobacillus mulieris Species Identifies Species-Specific Functionality

Catherine Putonti Loyola University Chicago, cputonti@luc.edu

Jason W. Shapiro Loyola University Chicago, jshapiro2@luc.edu

Adriana Ene Loyola University Chicago

Oleksandra Tsibere Loyola University Chicago

Alan J. Wolfe Loyola University Chicago, awolfe@luc.edu Follow this and additional works at: [https://ecommons.luc.edu/bioinformatics\\_facpub](https://ecommons.luc.edu/bioinformatics_facpub?utm_source=ecommons.luc.edu%2Fbioinformatics_facpub%2F56&utm_medium=PDF&utm_campaign=PDFCoverPages)

Part of the [Bioinformatics Commons,](https://network.bepress.com/hgg/discipline/110?utm_source=ecommons.luc.edu%2Fbioinformatics_facpub%2F56&utm_medium=PDF&utm_campaign=PDFCoverPages) and the [Biology Commons](https://network.bepress.com/hgg/discipline/41?utm_source=ecommons.luc.edu%2Fbioinformatics_facpub%2F56&utm_medium=PDF&utm_campaign=PDFCoverPages)

### Recommended Citation

Putonti, Catherine; Shapiro, Jason W.; Ene, Adriana; Tsibere, Oleksandra; and Wolfe, Alan J.. Comparative Genomic Study of Lactobacillus jensenii and the Newly Defined Lactobacillus mulieris Species Identifies Species-Specific Functionality. mSphere, 9, : , 2020. Retrieved from Loyola eCommons, Bioinformatics Faculty Publications, <http://dx.doi.org/10.1128/mSphere.00560-20>

This Article is brought to you for free and open access by the Faculty Publications and Other Works by Department at Loyola eCommons. It has been accepted for inclusion in Bioinformatics Faculty Publications by an authorized administrator of Loyola eCommons. For more information, please contact [ecommons@luc.edu.](mailto:ecommons@luc.edu)



This work is licensed under a [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/). © Putonti et al., 2020.





## **Comparative Genomic Study of Lactobacillus jensenii and the Newly Defined Lactobacillus mulieris Species Identifies Species-Specific Functionality**

**[Catherine Putonti,a](https://orcid.org/0000-0003-3049-5991),b,c [Jason W. Shapiro,a](https://orcid.org/0000-0002-1321-8418) Adriana Ene,b Oleksandra Tsibere,a [Alan J. Wolfec](https://orcid.org/0000-0003-4532-0545)**

aDepartment of Biology, Loyola University Chicago, Chicago, Illinois, USA <sup>b</sup>Bioinformatics Program, Loyola University Chicago, Chicago, Illinois, USA c Department of Microbiology and Immunology, Stritch School of Medicine, Loyola University Chicago, Maywood, Illinois, USA

**ABSTRACT** Lactobacilli are dominant members of the "healthy" female urogenital microbiota. One of these species, Lactobacillus jensenii, is routinely identified in the urinary microbiota of women both with and without urinary tract symptoms. In March 2020, the new bacterial species Lactobacillus mulieris was introduced, and phylogenetic and average nucleotide identity analysis identified eight L. jensenii strains that should be classified as members of the L. mulieris species. This prompted our phylogenomic study of all publicly available L. jensenii and L. mulieris genome sequences. While there is little variation in the 16S rRNA gene sequences, the core genome shows a clear distinction between genomes of the two species. We find eight additional strains of the species L. mulieris among these genomes. Furthermore, one strain, currently classified as L. mulieris UMB7784, is distinct from both L. jensenii and L. mulieris strains. As part of our comparative genomic study, we also investigated the genetic content that distinguishes these two species. Unique to the L. jensenii genomes are several genes related to catabolism of disaccharides. In contrast, L. mulieris genomes encode several cell surface and secreted proteins that are not found within the L. jensenii genomes. These L. jensenii-specific and L. mulierisspecific loci provide insight into phenotypic differences of these two species.

**IMPORTANCE** Lactobacillus species play a key role in the health of the urinary tract. For instance, Lactobacillus crispatus and L. jensenii have been found to inhibit uropathogenic Escherichia coli growth. While L. crispatus is typically found only within the microbiota of women without lower urinary tract symptoms (LUTS), L. jensenii has been found in the microbiota of women both with and without LUTS. With the recent introduction of the new species *Lactobacillus mulieris*, several strains of *L. jen*senii were reclassified as L. mulieris based upon gene marker and average nucleotide identity. We took a phylogenomic and comparative genomic approach to ascertain the genetic determinants of these two species. Looking at a larger data set, we identified additional L. mulieris strains, including one distinct from other members of the species—L. mulieris UMB7784. Furthermore, we identified unique loci in each species that may have clinical implications.

**KEYWORDS** Lactobacillus, Lactobacillus jensenii, Lactobacillus mulieris, urinary microbiome, urogenital microbiome

The Lactobacillus species L. crispatus, L. gasseri, L. iners, and L. jensenii are predominant members of the "healthy" female urogenital microbiota [\(1,](#page-5-0) [2\)](#page-5-1). L. jensenii has been shown to be a protective species, reducing growth of uropathogenic Escherichia coli and sexually transmitted infections [\(3](#page-5-2)[–](#page-5-3)[6\)](#page-5-4). Recently, Rocha et al. [\(7\)](#page-5-5) presented a new Lactobacillus species, L. mulieris. The type strain for this new species is L. mulieris

**Citation** Putonti C, Shapiro JW, Ene A, Tsibere O, Wolfe AJ. 2020. Comparative genomic study of Lactobacillus jensenii and the newly defined Lactobacillus mulieris species identifies speciesspecific functionality. mSphere 5:e00560-20. [https://doi.org/10.1128/mSphere.00560-20.](https://doi.org/10.1128/mSphere.00560-20) **Editor** Krishna Rao, University of Michigan— Ann Arbor

**Copyright** © 2020 Putonti et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0](https://creativecommons.org/licenses/by/4.0/) [International](https://creativecommons.org/licenses/by/4.0/) license.

Address correspondence to Catherine Putonti, [cputonti@luc.edu.](mailto:cputonti@luc.edu)

A deep dive into the genomes of L. jensenii and the new species L. mulieris finds unique loci that may have clinical implications. @PutontiLab

**Received** 10 June 2020 **Accepted** 27 July 2020 **Published** 12 August 2020







<span id="page-2-0"></span>FIG 1 Comparison of L. jensenii and L. mulieris genomes. (A) 16S rRNA gene sequence comparison. L. gasseri ATCC 33323 = JCM 1131 (GenBank accession no. [NC\\_008530\)](https://www.ncbi.nlm.nih.gov/nuccore/NC_008530) and L. crispatus ST1 [\(NC\\_014106\)](https://www.ncbi.nlm.nih.gov/nuccore/NC_014106) are included as an outgroup. The range of pairwise identity between groups is shown on the branches. (B) Phylogenetic tree of single-copy-number genes in the core genome. The range of pairwise identity between groups is shown on the branches. In the legend or key, the colors used for the species designation determined in this study and the isolation site are indicated for all genomes in both trees.

c10Ua161M, isolated from a urine sample. Rocha et al. [\(7\)](#page-5-5) found that the L. mulieris c10Ua161M genome, with eight L. jensenii strains, had an average nucleotide identity (ANI) with the L. jensenii type strain genome below the 95% species threshold [\(8\)](#page-5-6). Coinciding with the Rocha et al. publication [\(7\)](#page-5-5), we deposited and published two new Lactobacillus urinary isolates [\(9,](#page-5-7) [10\)](#page-5-8); while matrix-assisted laser desorption ionization– time of flight mass spectrometry (MALDI-TOF MS) classified these strains as L. jensenii, ANI analysis suggested they were members of the new L. mulieris species. This prompted us to investigate 43 publicly available L. jensenii and L. mulieris genomes (see Table S1 in the supplemental material).

First, we examined the L. jensenii and L. mulieris 16S rRNA gene sequences, which were  $\geq$ 99.54% identical [\(Fig. 1A\)](#page-2-0). The two species can be distinguished by just two nucleotides in the 16S rRNA gene sequence (see Fig. S1 in the supplemental material). The L. mulieris clade included the type strain, L. mulieris c10Ua161M, as well as strains identified by Rocha et al. [\(7\)](#page-5-5) as L. mulieris based upon pheS and rpoA gene sequence trees. Our two recent sequences UMB7784 and UMB9245, as well as six other strains, form a clade with the L. mulieris type strain, suggesting that they too represent this new species.

Next, the pangenome and set of single-copy genes in the core genome of the 43 L. jensenii and L. mulieris genomes were identified using the tool anvi'o [\(11\)](#page-5-9) (see Text S1 in the supplemental material). Phylogenomic analysis is based upon the concatenated protein sequences of the 453 single-copy core genes found, and sequence identities are reported for these concatenated sequences [\(Fig. 1B\)](#page-2-0). This tree showed clear distinction between genomes of the two species. Pairwise amino acid sequence comparisons between L. jensenii and L. mulieris core genomes ranged from 95.05 to 95.36% sequence identity. In contrast, the core genome sequence of the 26 genomes that form a clade with the L. jensenii type strain are 99.60% to 100% identical. On the basis of this core genome analysis, we propose that 17 strains should be classified as members of the species L. mulieris (Table S1).





<span id="page-3-0"></span>**FIG 2** Pangenome analysis of 26 L. jensenii (blue) and 17 L. mulieris (pink) genomes. Each ring corresponds to a single genome, and each radial extension in the ring corresponds to a particular gene. If the gene is present in the genome, the color is darker than if it is absent from the genome. The outer ring includes the percentage of the genomes including the gene. The three largest number of singleton genes from the center out are L. jensenii UMB0077, L. jensenii UMB8651, and L. mulieris UMB7784. The inset shows the increase in the pangenome size as new genomes are examined.

The variation within the L. mulieris clade of the core genome tree is primarily due to one isolate—L. mulieris UMB7784 [\(10\)](#page-5-8). The core genome of strain UMB7784 is less similar to other L. mulieris strains than they are to each other. Using JSpeciesWS [\(12\)](#page-5-10), we compared the two species' type strains to L. mulieris UMB7784. L. mulieris UMB7784 had an ANI value of 87.81% to L. jensenii DSM 20557 and 96.29% to L. mulieris c10Ua161M. Excluding UMB7784, the L. mulieris strains have an average ANI value of 99.66% (Table S2). While UMB7784 is best grouped with L. mulieris, additional sequencing of L. mulieris strains may reveal a third group to which it belongs.

The accessory genome for these strains includes 1,738 gene clusters [\(Fig. 2\)](#page-3-0), which is larger than the core genome. L. mulieris UMB7784 contains the most genes ( $n = 83$ ) that are unique to a single genome (singleton genes). Prior genome analyses of lactobacilli found that the genus has an open pangenome [\(13,](#page-5-11) [14\)](#page-5-12). Species that colonize diverse habitats and/or coexist with other microbes in large communities, such as many Lactobacillus species, typically have open pangenomes [\(15\)](#page-5-13). While the majority of comparative genome studies of lactobacilli have focused on those species most relevant to the dairy industry, a recent analysis of L. paragasseri and L. gasseri considered isolates from the human gut microbiota [\(16\)](#page-5-14). In addition to inhabiting the gut microbiota, L. paragasseri and L. gasseri are common inhabitants of the female urogenital tract, and Zhou et al. [\(16\)](#page-5-14) found that they also have an open pangenome. Our pangenome analysis of the L. jensenii and L. mulieris strains finds that they too have an open pangenome.

Last, we investigated the genetic content that distinguishes these two species. We examined each species individually and identified genes conserved among all genomes of that species that were not present in genomes of the other species. This analysis found 32 genes for L. jensenii (Table S3) and 62 genes for L. mulieris (Table S4). Some L. jensenii genes support the findings of Rocha et al. ([7\)](#page-5-5). For instance, L. jensenii can use ribose and trehalose, while L. mulieris cannot; only the L. jensenii genomes encode a ribose transporter and trehalose operon repressor (Table S3). Rocha et al. ( [7\)](#page-5-5) also noted that only L. jensenii can use arbutin. Our genome analysis identified a L. jensenii locus for disaccharide catabolism. Further investigation would be needed to ascertain whether this is arbutin. We also found new signs of functional specificity, including a conserved respiratory chain pathway present only in L. jensenii (Table S3) and multiple cell surface and secreted proteins in L. mulieris (Table S4). While none of these L. mulieris-specific genes are found in a  $L$ . jensenii strain, some of these genes do have homologs within L. psittaci, L. crispatus, and L. paragasseri (based on blastp to the nr database).

While other lactobacilli of the urinary tract have been associated with or without lower urinary tract symptoms (LUTS), L. jensenii has been found in communities regardless of urinary symptoms [\(17\)](#page-5-15). As our 16S rRNA gene sequence analysis suggests, distinguishing between L. jensenii and L. mulieris by 16S amplicon sequencing surveys is error-prone. Thus, whole-genome sequencing of the microbiota or directed searches for the genes identified here (Tables S3 and S4) is likely the best way to distinguish the two species in a community. With this knowledge, we can determine whether L. jensenii and/or L. mulieris have any association with LUTS. Furthermore, previous studies associating L. jensenii with the benefit of inhibiting the growth of urogenital pathogens must be revisited to assess whether both species have this beneficial role. The distinction of these two species may explain reported phenotypic variation among L. jensenii isolates, including  ${\sf H}_{\mathfrak{2}}{\sf O}_{\mathfrak{2}}$  production [\(18](#page-5-16), [19\)](#page-5-17). Twenty-three of the 43 genomes examined here are from our own collection of urinary isolates, and none of our 6 L. mulieris strains were isolated from women without LUTS (Table S1).

Recently, the entire Lactobacillus genus has been reevaluated given the availability of numerous complete and draft genome sequences [\(20](#page-5-18) , [21\)](#page-5-19). Our phylogenetic and phylogenomic analyses support the reclassification of the eight L. jensenii genomes identified by Rocha et al. ([7\)](#page-5-5) and eight additional genomes into the new L. mulieris species. Furthermore, we found that the L. mulieris UMB7784 genome is distinct from other L. mulieris strains. Looking at the 83 genes unique to this strain (Table S5), we found homologs in other lactobacilli, including L. salivarius , L. crispatus , L. hominis , L. johnsonii, and L. psittaci. UMB7784-specific genes include genes that encode the following metabolism-related proteins or pathways: glucose phosphotransferase system (PTS), arginase, glucocerebrosidase, diaminopropionate ammonia-lyase, and a phospholipase and a phosphate-selective porin. The UMB7784-specific genes also include genes encoding proteins related to survival: dTDP-4-dehydrorhamnose 3,5 epimerase, a bacteriocin, and a protein G-related albumin-binding molecule. L. mulieris UMB7784 suggests that the urogenital tract may contain additional lactobacilli subspecies or species. Further investigation of lactobacilli from the urogenital tract will provide critical insight into the genomic diversity of these two species and potential associations with urogenital symptoms and infections.

### **SUPPLEMENTAL MATERIAL**

Supplemental material is available online only. **TEXT S1**, DOCX file, 0.03 MB. **FIG S1**, PDF file, 0.1 MB.

**TABLE S1**, DOCX file, 0.02 MB. **TABLE S2**, DOCX file, 0.02 MB. **TABLE S3**, DOCX file, 0.01 MB. **TABLE S4**, DOCX file, 0.02 MB. **TABLE S5**, XLSX file, 0.01 MB.

#### <span id="page-5-0"></span>**REFERENCES**

- 1. Mendes-Soares H, Suzuki H, Hickey RJ, Forney LJ. 2014. Comparative functional genomics of Lactobacillus spp. reveals possible mechanisms for specialization of vaginal lactobacilli to their environment. J Bacteriol 196:1458 –1470. [https://doi.org/10.1128/JB.01439-13.](https://doi.org/10.1128/JB.01439-13)
- <span id="page-5-1"></span>2. Thomas-White K, Forster SC, Kumar N, Van Kuiken M, Putonti C, Stares MD, Hilt EE, Price TK, Wolfe AJ, Lawley TD. 2018. Culturing of female bladder bacteria reveals an interconnected urogenital microbiota. Nat Commun 9:1557. [https://doi.org/10.1038/s41467-018-03968-5.](https://doi.org/10.1038/s41467-018-03968-5)
- <span id="page-5-2"></span>3. Kalyoussef S, Nieves E, Dinerman E, Carpenter C, Shankar V, Oh J, Burd B, Angeletti RH, Buckheit KW, Fredricks DN, Madan RP, Keller MJ, Herold BC. 2012. Lactobacillus proteins are associated with the bactericidal activity against E. coli of female genital tract secretions. PLoS One 7:e49506. [https://doi.org/10.1371/journal.pone.0049506.](https://doi.org/10.1371/journal.pone.0049506)
- 4. Spurbeck RR, Arvidson CG. 2010. Lactobacillus jensenii surface-associated proteins inhibit Neisseria gonorrhoeae adherence to epithelial cells. Infect Immun 78:3103–3111. [https://doi.org/10.1128/IAI.01200-09.](https://doi.org/10.1128/IAI.01200-09)
- <span id="page-5-3"></span>5. Yamamoto HS, Xu Q, Fichorova RN. 2013. Homeostatic properties of Lactobacillus jensenii engineered as a live vaginal anti-HIV microbicide. BMC Microbiol 13:4. [https://doi.org/10.1186/1471-2180-13-4.](https://doi.org/10.1186/1471-2180-13-4)
- <span id="page-5-4"></span>6. Hudson PL, Hung KJ, Bergerat A, Mitchell C. 2020. Effect of vaginal Lactobacillus species on Escherichia coli growth. Female Pelvic Med Reconstr Surg 26:146 –151. [https://doi.org/10.1097/SPV.0000000000000827.](https://doi.org/10.1097/SPV.0000000000000827)
- <span id="page-5-5"></span>7. Rocha J, Botelho J, Ksiezarek M, Perovic SU, Machado M, Carriço JA, Pimentel LL, Salsinha S, Rodríguez-Alcalá LM, Pintado M, Ribeiro TG, Peixe L. 2020. Lactobacillus mulieris sp. nov., a new species of Lactobacillus delbrueckii group. Int J Syst Evol Microbiol 70:1522–1527. [https://](https://doi.org/10.1099/ijsem.0.003901) [doi.org/10.1099/ijsem.0.003901.](https://doi.org/10.1099/ijsem.0.003901)
- <span id="page-5-6"></span>8. Ramasamy D, Mishra AK, Lagier J-C, Padhmanabhan R, Rossi M, Sentausa E, Raoult D, Fournier P-E. 2014. A polyphasic strategy incorporating genomic data for the taxonomic description of novel bacterial species. Int J Syst Evol Microbiol 64:384 –391. [https://doi.org/10.1099/ijs.0.057091-0.](https://doi.org/10.1099/ijs.0.057091-0)
- <span id="page-5-7"></span>9. Ramotowski L, Miller-Ensminger T, Voukadinova A, Wolfe AJ, Putonti C. 2020. Draft genome sequence of Lactobacillus mulieris UMB9245, isolated from the female bladder. Microbiol Resour Announc 9:e00408-20. [https://doi.org/10.1128/MRA.00408-20.](https://doi.org/10.1128/MRA.00408-20)
- <span id="page-5-8"></span>10. Tsibere O, Miller-Ensminger T, Voukadinova A, Wolfe AJ, Putonti C. 2020. Draft genome sequence of Lactobacillus mulieris UMB7784, isolated from the female urinary tract. Microbiol Resour Announc 9:e00403-20. [https://doi.org/10.1128/MRA.00403-20.](https://doi.org/10.1128/MRA.00403-20)
- <span id="page-5-10"></span><span id="page-5-9"></span>11. Eren AM, Esen ÖC, Quince C, Vineis JH, Morrison HG, Sogin ML, Delmont TO. 2015. anvi'o: an advanced analysis and visualization platform for 'omics data. PeerJ 3:e1319. [https://doi.org/10.7717/peerj.1319.](https://doi.org/10.7717/peerj.1319)
- 12. Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc Natl Acad Sci U S A 106: 19126 –19131. [https://doi.org/10.1073/pnas.0906412106.](https://doi.org/10.1073/pnas.0906412106)
- <span id="page-5-11"></span>13. Sun Z, Harris HMB, McCann A, Guo C, Argimón S, Zhang W, Yang X, Jeffery IB, Cooney JC, Kagawa TF, Liu W, Song Y, Salvetti E, Wrobel A, Rasinkangas P, Parkhill J, Rea MC, O'Sullivan O, Ritari J, Douillard FP, Ross RP, Yang R, Briner AE, Felis GE, de Vos WM, Barrangou R, Klaenhammer TR, Caufield PW, Cui Y, Zhang H, O'Toole PW. 2015. Expanding the biotechnology potential of lactobacilli through comparative genomics of 213 strains and associated genera. Nat Commun 6:8322. [https://doi](https://doi.org/10.1038/ncomms9322) [.org/10.1038/ncomms9322.](https://doi.org/10.1038/ncomms9322)
- <span id="page-5-12"></span>14. Inglin RC, Meile L, Stevens MJA. 2018. Clustering of pan- and coregenome of Lactobacillus provides novel evolutionary insights for differentiation. BMC Genomics 19:284. [https://doi.org/10.1186/s12864-018](https://doi.org/10.1186/s12864-018-4601-5) [-4601-5.](https://doi.org/10.1186/s12864-018-4601-5)
- <span id="page-5-13"></span>15. Rouli L, Merhej V, Fournier P-E, Raoult D. 2015. The bacterial pangenome as a new tool for analysing pathogenic bacteria. New Microbes New Infect 7:72– 85. [https://doi.org/10.1016/j.nmni.2015.06.005.](https://doi.org/10.1016/j.nmni.2015.06.005)
- <span id="page-5-14"></span>16. Zhou X, Yang B, Stanton C, Ross RP, Zhao J, Zhang H, Chen W. 2020. Comparative analysis of Lactobacillus gasseri from Chinese subjects reveals a new species-level taxa. BMC Genomics 21:119. [https://doi.org/](https://doi.org/10.1186/s12864-020-6527-y) [10.1186/s12864-020-6527-y.](https://doi.org/10.1186/s12864-020-6527-y)
- <span id="page-5-15"></span>17. Pearce MM, Hilt EE, Rosenfeld AB, Zilliox MJ, Thomas-White K, Fok C, Kliethermes S, Schreckenberger PC, Brubaker L, Gai X, Wolfe AJ. 2014. The female urinary microbiome: a comparison of women with and without urgency urinary incontinence. mBio 5:e01283-14. [https://doi](https://doi.org/10.1128/mBio.01283-14) [.org/10.1128/mBio.01283-14.](https://doi.org/10.1128/mBio.01283-14)
- <span id="page-5-16"></span>18. Antonio MA, Hawes SE, Hillier SL. 1999. The identification of vaginal Lactobacillus species and the demographic and microbiologic characteristics of women colonized by these species. J Infect Dis 180: 1950 –1956. [https://doi.org/10.1086/315109.](https://doi.org/10.1086/315109)
- <span id="page-5-17"></span>19. Hütt P, Lapp E, Štšepetova J, Smidt I, Taelma H, Borovkova N, Oopkaup H, Ahelik A, Rööp T, Hoidmets D, Samuel K, Salumets A, Mändar R. 2016. Characterisation of probiotic properties in human vaginal lactobacilli strains. Microb Ecol Health Dis 27:30484. [https://doi.org/10.3402/mehd](https://doi.org/10.3402/mehd.v27.30484) [.v27.30484.](https://doi.org/10.3402/mehd.v27.30484)
- <span id="page-5-18"></span>20. Salvetti E, Harris HMB, Felis GE, O'Toole PW. 2018. Comparative genomics of the genus Lactobacillus reveals robust phylogroups that provide the basis for reclassification. Appl Environ Microbiol 84:e00993-18. [https://doi.org/10.1128/AEM.00993-18.](https://doi.org/10.1128/AEM.00993-18)
- <span id="page-5-19"></span>21. Zheng J, Wittouck S, Salvetti E, Franz CMAP, Harris HMB, Mattarelli P, O'Toole PW, Pot B, Vandamme P, Walter J, Watanabe K, Wuyts S, Felis GE, Gänzle MG, Lebeer S. 2020. A taxonomic note on the genus Lactobacillus: description of 23 novel genera, emended description of the genus Lactobacillus Beijerinck 1901, and union of Lactobacillaceae and Leuconostocaceae. Int J Syst Evol Microbiol 70:2782–2858. [https://doi.org/10.1099/](https://doi.org/10.1099/ijsem.0.004107) [ijsem.0.004107.](https://doi.org/10.1099/ijsem.0.004107)

