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5-28-2020

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Recommended Citation

Kalski, Martin; Miller-Ensminger, Taylor; Voukadinova, Adelina; Wolfe, Alan J.; and Putonti, Catherine. Draft Genome Sequence of Enterococcus faecalis UMB7780, Isolated from the Female Urinary Tract. Microbiology Resource Announcements, 9, : , 2020. Retrieved from Loyola eCommons, Bioinformatics Faculty Publications, <http://dx.doi.org/10.1128/MRA.00400-20>

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Microbiology AMERICAN SOCIETY FOR Resource Announcements MICROBIOLOGY

Draft Genome Sequence of Enterococcus faecalis UMB7780, Isolated from the Female Urinary Tract

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ABSTRACT Here, we present the draft genome sequence of Enterococcus faecalis UMB7780, isolated from the female urinary tract. The genome size is 3,005,901 bp, with a GC content of 37.36%, genome coverage of 179 \times , and an N_{50} score of 169,627 bp. Genome analysis identified evidence of antibiotic resistance, as well as intact prophages.

Enterococcus faecalis is part of the normal flora of the human microbiota, including
the gastrointestinal tract, oral cavity, and urinary tract (see review in reference [1\)](#page-2-0). E. faecalis has been found in the urinary tract microbiota (urobiome) of individuals with and thsoe without a lower urinary tract syndrome [\(2](#page-2-1)[–](#page-2-2)[6\)](#page-2-3). Enterococci are common nosocomial pathogens in humans [\(3](#page-2-4)[–](#page-2-5)[5\)](#page-2-2), and *E. faecalis* and *E. faecium* cause the majority of hospital-acquired enterococcal infections [\(6\)](#page-2-3). Likewise, E. faecalis is a common cause of urinary tract infections (UTI) [\(5,](#page-2-2) [7\)](#page-2-6). E. faecalis challenges medical institutes due to its ability to rapidly acquire resistance to a majority of antibiotics [\(6](#page-2-3)[–](#page-2-7)[9\)](#page-2-8). Here, we present the E. faecalis UMB7780 genome, collected from a catheterized urine sample obtained from a female patient with overactive bladder (OAB).

E. faecalis UMB7780 was isolated using the expanded quantitative urinary culture (EQUC) protocol [\(10\)](#page-2-9) from a prior institutional review board (IRB)-approved study (University of California, San Diego, IRB no. 170077AW). The genus and species for the isolate were determined by matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometry, following a protocol previously described [\(10,](#page-2-9) [11\)](#page-2-10), and the sample was stored at -80° C. From this freezer stock, the isolate was streaked onto a Columbia nalidixic acid (CNA) agar plate using aseptic techniques and incubated for 24 h at 35° C with 5% CO₂. A single colony was collected from the plate and cultured in liquid brain heart infusion (BHI) medium under the same conditions as described earlier. Genomic DNA was extracted using the Qiagen DNeasy blood and tissue kit. The kit's Gram-positive extraction protocol was modified as follows: 230 μ l of lysis buffer (180 μ l of 20 mM Tris-Cl, 2 mM sodium EDTA, and 1.2% Triton X-100 and 50 μ l of lysozyme) was used in step 2, and the incubation time in step 5 was reduced to 10 min. The extracted DNA was quantified using the Qubit fluorometer. The sample was sent to the Microbial Genome Sequencing Center (MiGS) at the University of Pittsburgh for sequencing. An Illumina tagmentation enzyme was used to fragment the DNA, and indices were attached using PCR. The library was then sequenced using the Illumina NextSeq 550 platform, producing 2,197,539 pairs of 150-bp reads. The raw reads were trimmed using Sickle v1.33 [\(https://github.com/najoshi/sickle\)](https://github.com/najoshi/sickle) and assembled using SPAdes v3.13.0 with the "only-assembler" option for k values of 55, 77, 99, and 127 [\(12\)](#page-2-11). Genome coverage was calculated using BBMap v38.76 [\(https://sourceforge](https://sourceforge.net/projects/bbmap/) [.net/projects/bbmap/\)](https://sourceforge.net/projects/bbmap/). The genome was initially annotated using PATRIC v3.6.3 [\(13\)](#page-2-12)

Citation Kalski M, Miller-Ensminger T, Voukadinova A, Wolfe AJ, Putonti C. 2020. Draft genome sequence of Enterococcus faecalis UMB7780, isolated from the female urinary tract. Microbiol Resour Announc 9:e00400-20. [https://doi.org/10.1128/MRA.00400-20.](https://doi.org/10.1128/MRA.00400-20)

Editor Vincent Bruno, University of Maryland School of Medicine

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Received 13 April 2020 **Accepted** 1 May 2020 **Published** 28 May 2020 but then reannotated with the Prokaryotic Genome Annotation Pipeline (PGAP) v4.11 [\(14\)](#page-2-13). PGAP annotation was used for the publicly available assembly. Unless otherwise mentioned, default parameters were used for all software tools.

The E. faecalis UMB7780 draft genome is 3,005,901 bp and assembled into 61 contigs with a genome coverage of 179 \times and an N_{50} score of 169,627 bp. The GC content is 37.36%. The PGAP annotation identified 2,802 protein-coding genes, 51 tRNAs, and 3 rRNA operons. The PATRIC annotation identified 47 genes associated with antimicrobial resistance. The ResFinder v 3.2 server predicts that UMB7780 is resistant to macrolides and tetracycline [\(15\)](#page-2-14). The website PHASTER [\(16\)](#page-2-15) identified 2 incomplete and 2 intact prophages within the strain's genome. One CRISPR array was detected by the CRISPRCasFinder website [\(17\)](#page-2-16). Additional testing and analysis of this strain will provide a better understanding of antibiotic resistance within the human urobiome.

Data availability. This whole-genome shotgun project has been deposited in GenBank under the accession no. [JAAUVY000000000.](https://www.ncbi.nlm.nih.gov/nuccore/JAAUVY000000000) The version described in this paper is the first version, JAAUVY010000000. The raw sequencing reads have been deposited in the SRA under the accession no. [SRR11441029.](https://www.ncbi.nlm.nih.gov/sra/SRR11441029)

ACKNOWLEDGMENTS

This work was conducted as part of the Bacterial Genomics course at Loyola University Chicago's Department of Biology. For prior patient recruitment, we acknowledge the Loyola Urinary Education and Research Collaborative (LUEREC) and the patients who provided the samples for this study.

REFERENCES

- 1. Kao PHN, Kline KA. 2019. Dr. Jekyll and Mr. Hide: how Enterococcus faecalis subverts the host immune response to cause infection. J Mol Biol 431:2932–2945. [https://doi.org/10.1016/j.jmb.2019.05.030.](https://doi.org/10.1016/j.jmb.2019.05.030)
- 2. Horsley H, Malone-Lee J, Holland D, Tuz M, Hibbert A, Kelsey M, Kupelian A, Rohn JL. 2013. Enterococcus faecalis subverts and invades the host urothelium in patients with chronic urinary tract infection. PLoS One 8:e83637. [https://doi.org/10.1371/journal.pone.0083637.](https://doi.org/10.1371/journal.pone.0083637)
- 3. Fiore E, Van Tyne D, Gilmore MS. 2019. Pathogenicity of enterococci. Microbiol Spectr 7:GPP3-0053-2018. [https://doi.org/10.1128/microbiolspec](https://doi.org/10.1128/microbiolspec.GPP3-0053-2018) [.GPP3-0053-2018.](https://doi.org/10.1128/microbiolspec.GPP3-0053-2018)
- 4. Gordon S, Swenson JM, Hill BC, Pigott NE, Facklam RR, Cooksey RC, Thornsberry C, Jarvis WR, Tenover FC. 1992. Antimicrobial susceptibility patterns of common and unusual species of enterococci causing infections in the United States. Enterococcal Study Group. J Clin Microbiol 30:2373–2378. [https://doi.org/10.1128/JCM.30.9.2373-2378.1992.](https://doi.org/10.1128/JCM.30.9.2373-2378.1992)
- 5. Kau AL, Martin SM, Lyon W, Hayes E, Caparon MG, Hultgren SJ. 2005. Enterococcus faecalis tropism for the kidneys in the urinary tract of C57BL/6J mice. Infect Immun 73:2461–2468. [https://doi.org/10.1128/IAI](https://doi.org/10.1128/IAI.73.4.2461-2468.2005) [.73.4.2461-2468.2005.](https://doi.org/10.1128/IAI.73.4.2461-2468.2005)
- 6. Guzman Prieto AM, van Schaik W, Rogers MRC, Coque TM, Baquero F, Corander J, Willems RJL. 2016. Global emergence and dissemination of enterococci as nosocomial pathogens: attack of the clones? Front Microbiol 7:788. [https://doi.org/10.3389/fmicb.2016.00788.](https://doi.org/10.3389/fmicb.2016.00788)
- 7. Gawryszewska I, Malinowska K, Kuch A, Chrobak-Chmiel D, Trokenheim ŁŁ, Hryniewicz W, Sadowy E. 2017. Distribution of antimicrobial resistance determinants, virulence-associated factors and clustered regularly interspaced palindromic repeats loci in isolates of Enterococcus faecalis from various settings and genetic lineages. Pathog Dis 75:ftx021. [https://](https://doi.org/10.1093/femspd/ftx021) [doi.org/10.1093/femspd/ftx021.](https://doi.org/10.1093/femspd/ftx021)
- 8. Miller WR, Munita JM, Arias CA. 2014. Mechanisms of antibiotic resistance in enterococci. Expert Rev Anti Infect Ther 12:1221–1236. [https://](https://doi.org/10.1586/14787210.2014.956092) [doi.org/10.1586/14787210.2014.956092.](https://doi.org/10.1586/14787210.2014.956092)
- 9. Hollenbeck BL, Rice LB. 2012. Intrinsic and acquired resistance mechanisms in enterococcus. Virulence 3:421– 433. [https://doi.org/10.4161/viru](https://doi.org/10.4161/viru.21282) [.21282.](https://doi.org/10.4161/viru.21282)
- 10. Hilt EE, McKinley K, Pearce MM, Rosenfeld AB, Zilliox MJ, Mueller ER, Brubaker L, Gai X, Wolfe AJ, Schreckenberger PC. 2014. Urine is not sterile: use of enhanced urine culture techniques to detect resident bacterial flora in the adult female bladder. J Clin Microbiol 52:871– 876. [https://doi.org/10.1128/JCM.02876-13.](https://doi.org/10.1128/JCM.02876-13)
- 11. Price TK, Dune T, Hilt EE, Thomas-White KJ, Kliethermes S, Brincat C, Brubaker L, Wolfe AJ, Mueller ER, Schreckenberger PC. 2016. The clinical urine culture: enhanced techniques improve detection of clinically relevant microorganisms. J Clin Microbiol 54:1216 –1222. [https://doi.org/](https://doi.org/10.1128/JCM.00044-16) [10.1128/JCM.00044-16.](https://doi.org/10.1128/JCM.00044-16)
- 12. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455– 477. [https://doi.org/10.1089/cmb.2012.0021.](https://doi.org/10.1089/cmb.2012.0021)
- 13. Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. [https://](https://doi.org/10.1038/srep08365) [doi.org/10.1038/srep08365.](https://doi.org/10.1038/srep08365)
- 14. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614 – 6624. [https://doi.org/10.1093/nar/gkw569.](https://doi.org/10.1093/nar/gkw569)
- 15. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640 –2644. [https://doi](https://doi.org/10.1093/jac/dks261) [.org/10.1093/jac/dks261.](https://doi.org/10.1093/jac/dks261)
- 16. Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. Nucleic Acids Res 44:W16 –W21. [https://doi.org/10.1093/nar/gkw387.](https://doi.org/10.1093/nar/gkw387)
- 17. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a Web tool to identify clustered regularly interspaced short palindromic repeats. Nucleic Acids Res 35:W52–W57. [https://doi.org/10.1093/nar/gkm360.](https://doi.org/10.1093/nar/gkm360)