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Draft Genome Sequence of *Enterobacter asburiae* UFMG-H9, Isolated from Urine from a Healthy Bovine Heifer (Gyr Breed)

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ABSTRACT *Enterobacter asburiae* is part of the *Enterobacter cloacae* complex, related to nosocomial opportunistic infections in humans. Here, we report the draft genome of *E. asburiae* strain UFMG-H9, an isolate from urine from a healthy Gyr heifer.

Enterobacter asburiae is part of the *Enterobacter cloacae* complex, which includes six species of the genus. Species of this complex are widely distributed and known as commensal species and opportunistic pathogens with the potential to cause nosocomial infections in humans (1). *E. asburiae* has also been identified as an antibiotic-resistant species (2), and although it has not been reported in livestock animals so far, it was recently reported in fruits and vegetables (3), increasing concerns about the dissemination of this pathogen. Here, we announce the draft genome sequence of *E. asburiae* strain UFMG-H9. This strain was isolated from a healthy heifer in Brazil. The animal belongs to a pure-by-origin Gyr herd, from the Agricultural Research Company of Minas Gerais state.

The present study was approved by the Ethics Committee in Animal Experimentation of the Universidade Federal de Minas Gerais, Brazil (approval number 40/2019), and samples were collected in May 2019, with all of the experiments being performed in accordance with relevant guidelines. For sampling, the vulva of the animal was washed with soap and distilled water, and then midstream urine collection was made using a 50-ml sterile tube. The material was frozen and kept at -20°C until processing (within 48 h). Two-milliliter aliquots were centrifuged, and the supernatant was plated onto lysogeny broth (LB) agar plates. The plates were incubated overnight at 37°C . Single colonies were selected, grown in LB cultures overnight at 37°C , and plated again. In order to obtain pure colonies, this process was repeated at least three times. A single colony was then inoculated in LB liquid medium and grown overnight at 37°C with agitation. A 1-ml aliquot was centrifuged, and the pellet was used for DNA extraction using the DNeasy UltraClean microbial kit (Qiagen, Hilden, Germany). The genus and species were determined by sequencing of the 16S rRNA gene using the 63f and 1387r primers (4). The amplicons were sequenced by Genewiz (South Plainfield, NJ, USA) via Sanger sequencing, and the sequence produced was queried against the NCBI 16S rRNA sequence database. DNA was quantified with a Qubit fluorometer and sent to the Microbial Genomic Sequencing Center at the University of Pittsburgh for whole-genome sequencing. To prepare the library, the DNA was fragmented using an Illumina tagmentation enzyme, and indices were attached using PCR. The genome was sequenced using the NextSeq 550 platform. Sequencing produced 1,406,751 pairs of

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TABLE 1 Predicted antibiotic resistances for *E. asburiae* UFMG-H9

Resistance gene	Predicted phenotype
<i>bla</i> _{ACT-6}	β-Lactam resistance (AmpC type)
<i>fosA</i>	Fosfomycin resistance
<i>qnrB2</i>	Quinolone resistance
<i>sul1</i>	Sulfonamide resistance
<i>dfrA25</i>	Trimethoprim resistance

150-bp reads, which were trimmed using Sickle v1.33 (<https://github.com/najoshi/sickle>). Once trimmed, the genome was assembled using SPAdes v3.13.0 with the only-assembler option for k values of 55, 77, 99, and 127 (5). The genome coverage was then calculated using BBDMap v38.47 (<https://sourceforge.net/projects/bbmap>). After that, the genome was annotated by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v4.11 (6). Unless noted otherwise, default parameters were used for all software.

The *E. asburiae* UFMG-H9 draft genome is 5,039,202 bp long with a GC content of 52.71%. The assembly includes 116 contigs with an N_{50} value of 139,398 bp, and the genome coverage was 77×. Annotation identified 4,765 coding genes and 77 tRNAs. The genome sequence was examined using ResFinder v3.2 (7), which revealed several antibiotic resistances (Table 1). Future analysis regarding the pathogenic potential of this strain would further our knowledge of *E. asburiae* circulation.

Data availability. This whole-genome sequencing project is available in GenBank under the accession number [JAAVSG000000000](https://www.ncbi.nlm.nih.gov/nuclseq/JAAVSG000000000). Raw reads were deposited in the SRA under accession number [SRR11455637](https://www.ncbi.nlm.nih.gov/sra/SRR11455637). This sequencing effort is part of BioProject accession number [PRJNA615899](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA615899).

REFERENCES

1. Jiménez-Guerra G, Borrego-Jiménez J, Gutiérrez-Soto B, Expósito-Ruiz M, Navarro-Marí JM, Gutiérrez-Fernández J. 2020. Susceptibility evolution to antibiotics of *Enterobacter cloacae*, *Morganella morganii*, *Klebsiella aerogenes* and *Citrobacter freundii* involved in urinary tract infections: an 11-year epidemiological surveillance study. *Enferm Infecc Microbiol Clin* 38:166–169. <https://doi.org/10.1016/j.eimc.2019.07.010>.
2. Oshiro S, Tada T, Watanabe S, Tohya M, Hishinuma T, Uchida H, Kuwahara-Arai K, Mya S, Zan KN, Kirikae T, Tin HH. 2020. Emergence and spread of carbapenem-resistant and aminoglycoside-panresistant *Enterobacter cloacae* complex isolates coproducing NDM-type metallo-β-lactamase and 16S rRNA methylase in Myanmar. *mSphere* 5:e00054-20. <https://doi.org/10.1128/mSphere.00054-20>.
3. Al-Kharousi ZS, Guizani N, Al-Sadi AM, Al-Bulushi IM. 2019. Antibiotic resistance of *Enterobacteriaceae* isolated from fresh fruits and vegetables and characterization of their AmpC β-lactamases. *J Food Prot* 82: 1857–1863. <https://doi.org/10.4315/0362-028X.JFP-19-089>.
4. Marchesi JR, Sato T, Weightman AJ, Martin TA, Fry JC, Hiom SJ, Dymock D, Wade WG. 1998. Design and evaluation of useful bacterium-specific PCR primers that amplify genes coding for bacterial 16S rRNA. *Appl Environ Microbiol* 64:795–799. <https://doi.org/10.1128/AEM.64.2.795-799.1998>.
5. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
6. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44: 6614–6624. <https://doi.org/10.1093/nar/gkw569>.
7. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother* 67:2640–2644. <https://doi.org/10.1093/jac/dks261>.