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Interspecies variation in hominid gut microbiota controls host gene regulation

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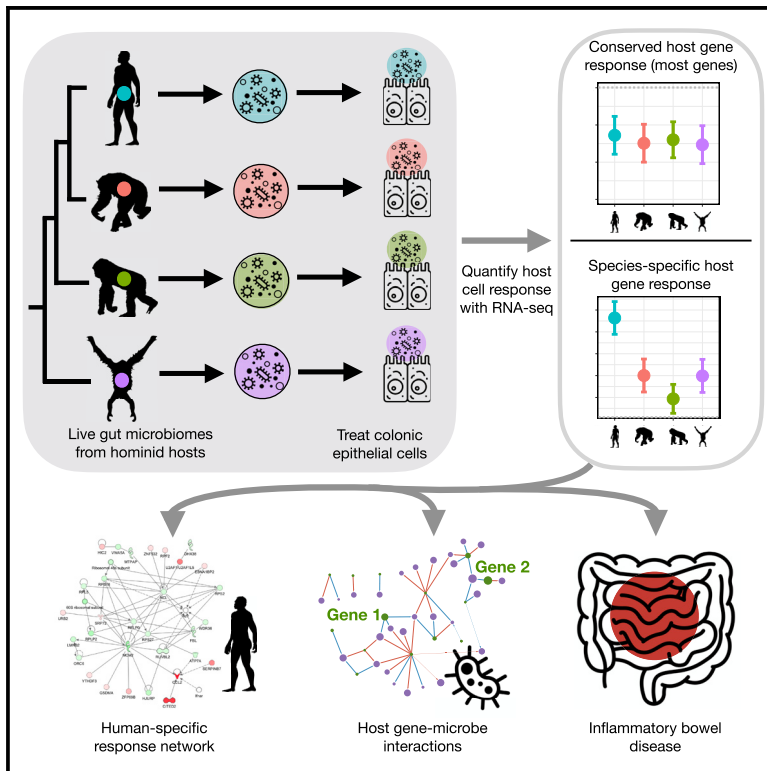
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Graphical abstract



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In brief

Muehlbauer et al. investigate how variation between different hominid microbiomes drives host gene expression in colonic epithelial cell cultures. They find that host genes that respond only to microbiomes from a specific hominid species are linked to gastrointestinal diseases, suggesting implications for understanding how the microbiome can impact human health.

Highlights

- Gut microbiota from humans, chimpanzees, gorillas, and orangutans regulate host genes
- Most host genes respond to all hominid microbiomes in the same way
- Some host genes respond only to microbiomes from a specific hominid species
- Genes with such species-specific response are linked to human intestinal diseases



Resource

Interspecies variation in hominid gut microbiota controls host gene regulation

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SUMMARY

The gut microbiome exhibits extreme compositional variation between hominid hosts. However, it is unclear how this variation impacts host physiology across species and whether this effect can be mediated through microbial regulation of host gene expression in interacting epithelial cells. Here, we characterize the transcriptional response of human colonic epithelial cells *in vitro* to live microbial communities extracted from humans, chimpanzees, gorillas, and orangutans. We find that most host genes exhibit a conserved response, whereby they respond similarly to the four hominid microbiomes. However, hundreds of host genes exhibit a divergent response, whereby they respond only to microbiomes from specific host species. Such genes are associated with intestinal diseases in humans, including inflammatory bowel disease and Crohn's disease. Last, we find that inflammation-associated microbial species regulate the expression of host genes previously associated with inflammatory bowel disease, suggesting health-related consequences for species-specific host-microbiome interactions across hominids.

INTRODUCTION

The microbiome of the primate gastrointestinal tract plays an important role in host physiology and health. Extreme variation in the gut microbiome has been observed between healthy human individuals; this variation is even more pronounced between different species of great apes (Human Microbiome Project Consortium, 2012; Nishida and Ochman, 2019). Microbiome composition is strongly correlated with the species of the host, a pattern known as co-diversification. Within hominids and other nonhuman primates, co-diversification between host and microbial symbionts has led to overall microbiome composition clustering along the expected phylogenetic relationships of the host species, including bacterial, archeal, and eukaryotic groups within the gut microbiome (Ochman et al., 2010; Moeller et al., 2012; Raymann et al., 2017; Mann et al., 2020; Amato et al., 2019a). However, reports show that these phylogenetic constraints are flexible, depending on diet and subsistence strategy

(Gomez et al., 2019). For example, compared with industrialized human groups, small scale rural or agricultural human populations share a greater number of gut microbiome traits with wild nonhuman primates (Gomez et al., 2019; Amato et al., 2019b).

Different hominid species harbor many of the same bacterial phyla in the gastrointestinal tract, but in varying abundances. For example, both the human and chimpanzee guts are primarily colonized by Bacteroidetes and Firmicutes, but the chimpanzee gut also harbors higher abundances of microbial phyla that are relatively rare in humans, including Actinobacteria, Euryarchaeota, Tenericutes, and Verrucomicrobia (Ochman et al., 2010; Nishida and Ochman, 2019). Gorillas, besides also displaying the presence of these rare taxa, harbor greater abundances of Chloroflexi, Tenericutes, and Fibrobacteres (Gomez et al., 2015, 2016b; Hicks et al., 2018). Although the orangutan microbiome has not been characterized as thoroughly, a previous report has shown that orangutan guts harbor higher diversity in archaeal lineages compared to other great apes, in addition to



similar microbial phyla as gorillas and chimpanzees (Raymann et al., 2017; Delsuc et al., 2014). At lower microbial taxonomic levels, very different microbial species are present in human and chimpanzee microbiomes, resulting in greater divergence (Nishida and Ochman, 2019).

Overall gut microbiome composition is shaped by a combination of host genetics, host physiology, and environmental factors. Studies have shown that host genetic variation influences microbiome composition within humans, but has yet to be studied in other hominids (Blekhnman et al., 2015; Goodrich et al., 2014). Among environmental influences, diet has a large impact on the primate gut microbiome (Nagpal et al., 2018; Hicks et al., 2018; Gomez et al., 2016b). Most non-human great ape species in the wild and in captivity subsist on a primarily plant-based diet of fruit and vegetation that is occasionally supplemented by animal protein, such as meat or insects (Vogel et al., 2015; Tutin and Fernandez, 1993; Watts et al., 2012). In contrast, human diets are usually omnivorous and highly variable depending on cultural influences, agricultural practices, geographic location, and individual dietary preferences (Wu et al., 2011; Lang et al., 2018). Other environmental factors that can influence microbiome composition between primates include variation in geography, seasonality, and other social behaviors such as grooming (Tung et al., 2015; Grieneisen et al., 2019). In addition, physiological differences between primate species, such as differences in gut morphology and digestive processes, also contribute to differences in microbiome composition (Amato et al., 2019a). Although a large effort has been made to characterize the factors that influence variation in the microbiome, it is unclear how variation in microbiome composition between great ape species can impact relevant host phenotypes.

A likely mechanism by which the microbiome can affect host physiology is through regulating the expression of host genes in interacting intestinal epithelial cells (Luca et al., 2018; Richards et al., 2016, 2019). Studies in animal models have demonstrated that gut microbiota can drive changes in host gene expression by altering epigenetic programming, such as histone modification, transcription factor binding, and methylation (Qin et al., 2018; Camp et al., 2014; Pan et al., 2018; Krautkrämer et al., 2016). For example, Camp et al. (2014) found that the microbiome drives the differential expression of transcription factors enriched in accessible binding sites. In addition, Pan et al. (2018) found that the microbiome can alter DNA methylation in the gut epithelial cells of mice. Moreover, in cell culture, inter-individual variation in microbiome composition can drive differential responses in host gene expression at the intestinal level (Richards et al., 2019). However, we do not know how interspecies variation in the microbiome affects gene regulation in host cells. When considering microbiota variation among great ape species and their influences on host gene expression, *in vivo* studies in experimental animal models are limited. Furthermore, *in vivo* experiments can be confounded by a multitude of factors, such as differences in diet between the animal model species and the primate species of interest, microbiota colonization history of the animal model, and inherent differences in the genetic backgrounds between the animal model and the primate species (Luca et al., 2018).

Here, we use an *in vitro* experimental system (Richards et al., 2019, 2016) to assess host gene expression changes in response to diverse gut microbiota from four great ape species: humans (*Homo sapiens*), and captive chimpanzees (*Pan troglodytes*), gorillas (*Gorilla gorilla gorilla*), and orangutans (*Pongo abelii*). We collected microbiomes from four humans, three chimpanzees, six gorillas, and three orangutans as biological replicates. In each of the non-human primate groups, one of the individuals donated two samples, bringing the total number of samples to four human, four chimpanzee, seven gorilla, and four orangutan samples. This experimental design allows us to determine causal relationships between gut microbiome composition and gene expression changes in colonic epithelial cells that are induced by the microbiome while controlling for potentially confounding environmental and technical effects (Richards et al., 2016, 2019). We have leveraged this design to ascertain how host genes respond to between-species variation in microbiome composition across hominids, characterize the function of host genes that respond to microbiota from each great ape species, and identify microbial taxa and pathways that likely drive expression of specific host genes.

RESULTS

To assess how host genes respond to variation in the microbiome, we extracted live microbiota from 19 fecal samples from 16 individuals from four hominid species (4 humans, 3 chimpanzees, 6 gorillas, and 3 orangutans), and treated human colonic epithelial cells (colonocytes) with the extracted microbiota using an experimental technique from a previously published method (Table S1) (Richards et al., 2016, 2019). Briefly, colonocytes are treated with live primate microbiota for 2 h, after which we quantify changes in gene expression in the colonocytes using RNA sequencing (RNA-seq) (Figure 1A; see STAR Methods). Additionally, we used 16S rRNA sequencing and shotgun metagenomics to characterize the composition of the microbiome in these samples. A principal coordinate analysis of Bray-Curtis dissimilarities shows that the microbiome samples do not change during the treatment, due to the short culturing time. Importantly, microbiome samples cluster by primate host species of origin (Figures 1B and S1A). This observation is consistent with previous findings showing that the phylogenetic relationship between primate host species is reflected in their microbiomes (Ochman et al., 2010), and indicates that interspecies microbiome distinctions between wild apes is maintained in the captive individuals included in our study. We note that in a comparison of our captive chimpanzees and gorillas samples with publicly available data from wild chimpanzees and gorillas (Campbell et al., 2020), we find that, as expected, captive microbiomes differ from their wild counterparts (Figure S1B). However, the clear clustering of the microbiomes used in our study by species (Figure 1B) indicates that strong inter-species distinctions in the gut microbiome persist in these captive individuals.

The bacterial composition of the samples confirmed clear distinctions between hominid species at the phylum level (Figures 1C and S1C), with nine of the most abundant microbial phyla showing significantly different levels between hominid species

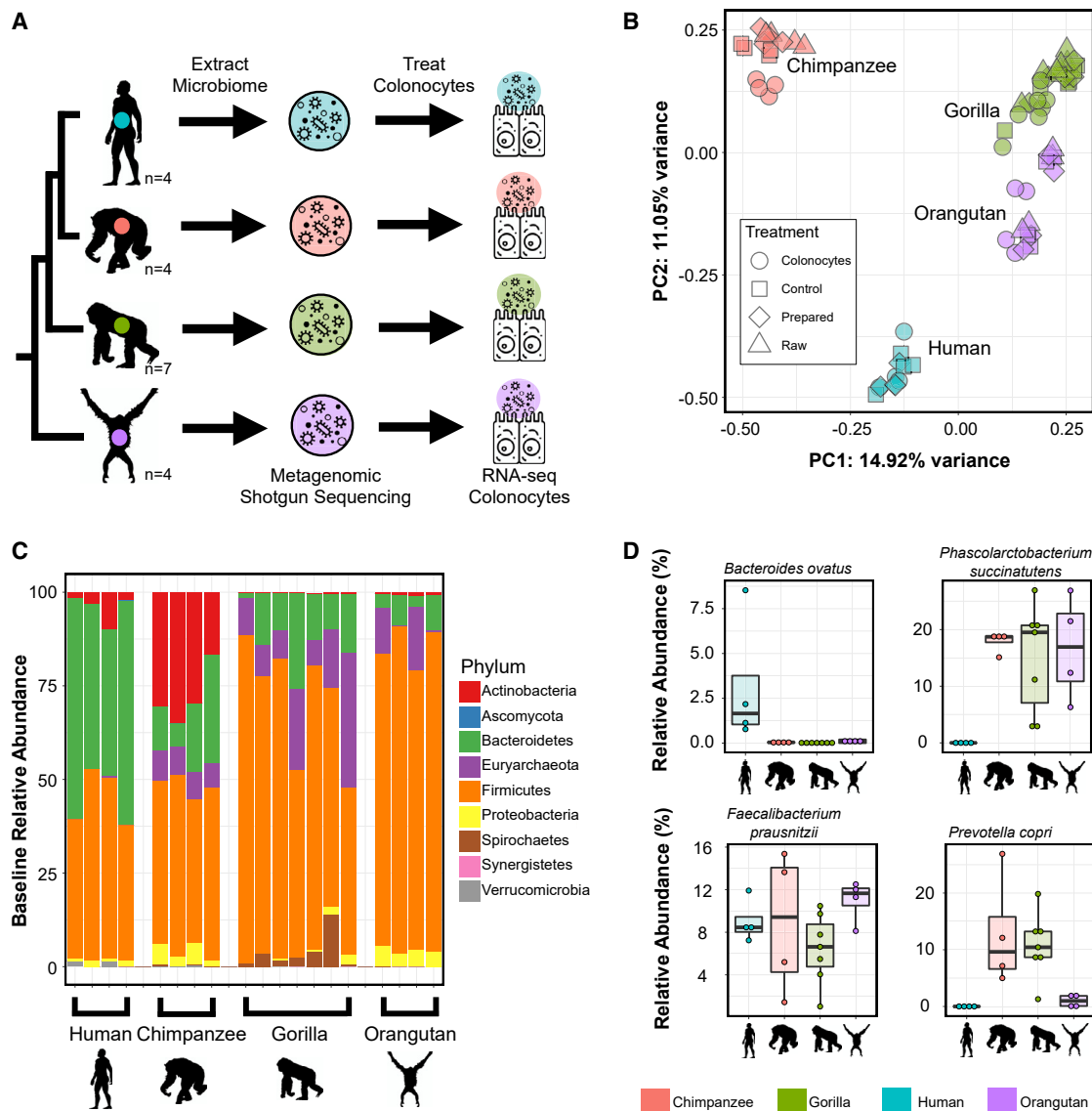


Figure 1. Experimental design and gut microbiome composition

(A) Schematic of the experimental design. Live microbiomes were extracted from fecal samples from humans (n = 4, blue) chimpanzees (n = 4, orange), gorillas (n = 7, green), and orangutans (n = 4; purple), and were sequenced with metagenomic shotgun sequencing. Microbes were incubated with human colonic epithelial cells for 2 h, after which host response was profiled through RNA-seq on the epithelial cells, and the microbiome was profiled via 16S rRNA sequencing. See also [Table S1](#).

(B) PCOA plot showing the Bray-Curtis dissimilarity of all the microbiome samples from all four primate species (represented by the same colors in A) at different stages of the experiment. Here, “Raw” refers to the microbiomes of raw fecal samples; “Prepared” refers to microbiome samples that have been prepared (see [STAR Methods](#)) but have not been cultured with colonocytes; “Colonocytes” refers to microbiomes after incubation with colonocytes; “Control” refers to microbiomes that have been incubated without colonocytes. See also [Figure S1A](#).

(C) Relative abundances of microbial phyla from shotgun metagenomic sequencing for each hominid fecal sample, sequenced before being cultured with colonocytes. The legend on the right indicates the colors corresponding to each phylum. See also [Figures S1B](#) and [S1C](#).

(D) Examples of microbial species (from shotgun metagenomics sequencing) with various patterns of abundance across hominid species. In each panel, the x axis indicates the primate host species, whereas the y axis indicates the relative abundance of the microbial species. Each dot represents the abundance of the microbial species in a hominid individual host. *Bacteroides ovatus* (top left) shows a high abundance in humans relative to the other hominid species. *Phascolarctobacterium succinatutens* is highly abundant in the non-human hominids but not present in the human microbiomes. *Faecalibacterium prausnitzii* is highly abundant in all four hominid species. *Prevotella copri* is highly abundant in chimpanzees and gorillas, has a lower abundance in orangutans, and is not present in the human samples. See also [Figure S1C](#) and [Table S2](#).

(Table S2; Kruskal-Wallis test, Benjamini-Hochberg false discovery rate [FDR] <0.1). The human microbiome samples have a high relative abundance of Bacteroidetes and Firmicutes, which have both been previously identified as dominant phyla in the human gut (Human Microbiome Project Consortium, 2012; Turnbaugh et al., 2007). In addition, Actinobacteria abundance is significantly different between hominid species (Kruskal-Wallis test, Benjamini-Hochberg q-value = 0.00567; ANOVA, Benjamini-Hochberg q-value = 3.82×10^{-9}), with chimpanzees showing the greatest abundance (see Figure 1C). Furthermore, we identified 21 microbial species that are differentially abundant between hominid host species (Table S2; Kruskal-Wallis, Benjamini-Hochberg FDR <0.1). Examples of several microbes that have variable abundance across species, including *Bacteroides ovatus*, which shows higher abundance in humans compared to other hominids; *Phascolarctobacterium succinatutens*, which shows lower abundance in humans compared to other hominids; and *Prevotella copri*, which has higher abundance in gorilla and orangutan, are shown in Figure 1D.

To characterize the host response to the microbiome, we used likelihood ratio tests combined with a negative binomial model (DESeq2) to identify host genes that change their expression after inoculation with microbiomes from the four hominid host species (see STAR Methods). We identified 4,329 host genes that respond to the microbiome of at least one hominid species (Figure 2A, Benjamini-Hochberg FDR <0.1). The majority of differentially expressed genes (2,261 genes, 52%) respond to the microbiomes of all four hominids (Figures 2A and 2B; Table S3, full dataset available; STAR Methods). Despite this overall consistent response, we find 164 host genes that respond in a species-specific manner; namely, respond to the microbiome of one hominid species but not the other three. For example, *SHROOM3* responds to the human microbiome, but shows no response to the chimpanzee, gorilla, and orangutan microbiomes (Figure 2C). Similarly, *B3GAT2*, *DUSP11*, and *DARS2* respond in a species-specific manner to the chimpanzee, gorilla, and orangutan microbiomes, respectively (Figure 2C). We also find 394 host genes that respond to microbiomes from two hominid species (e.g., *CBR1* responds to orangutan and gorilla microbiomes) (Figure 2C). Likewise, 1,313 host genes respond to microbiomes from three hominid species, and 13,531 genes show no response to any of the hominid microbiomes (e.g., *INVS*) (Figure 2C).

To understand how genes with a host species-specific response may interact with each other, we visualized interaction networks for differentially expressed host genes that respond to microbiomes from each hominid species (Kr amer et al., 2014) (Ingenuity Pathway Analysis, <https://www.qiagenbioinformatics.com>) (Figures 3A, 3B, S2A, and S2B; STAR Methods). The most significant interaction network of host genes that respond only to human microbiomes is enriched with functional categories related to cancer, cell death and survival, and organismal injuries and abnormalities (Figure 3A; Table S4). This is consistent with previous studies showing that the microbiome may influence host disease through changes in host gene regulation, but also suggests that this effect may be specific to human microbiomes (Qin et al., 2018; Camp et al., 2014; Pan et al., 2018; Krautkramer et al., 2016). By comparison, the most significant interaction

network of genes that respond specifically to orangutan microbiomes is enriched for functional categories related to carbohydrate metabolism, lipid metabolism, and small molecule biochemistry (Figure 3B; Table S4). This is consistent with the observation that orangutan diets, compared to that of gorillas or chimpanzees, could incorporate a greater proportion of ripe fruits and highly digestible/simple sugars in peak seasons (up to 100% dependence on fruit) (Remis, 1997; Taylor, 2006). In addition, previous reports point to a highly diverse archaeal community in orangutans compared to other apes, which could be associated with an increased capacity to metabolize highly fermentable plant materials (Raymann et al., 2017). See Table S4 for functions enriched in the most significant networks for genes that respond only to gorilla microbiomes and only to chimpanzee microbiomes.

To further characterize the biological functions represented by host genes that respond to variation in hominid microbiomes, we categorized differentially expressed genes into two groups: low-divergence genes, which show a similar magnitude and direction of response to the four hominid microbiomes, and high-divergence genes, which show a highly variable response to the four hominid microbiomes (following the approach of Hagai et al. [2018]) (Table S5; STAR Methods). We find that low-divergence genes, namely, differentially expressed genes that show a similar response to microbiomes from all four primate species, tend to be enriched for functions related to basic cell processes, such as RNA processing, cell cycle, and RNA metabolic processing (Figure 3C, Benjamini-Hochberg FDR <0.1; Table S6). This suggests that these genes are likely involved in basic host responses to bacterial cells, rather than response to specific microbial features. Interestingly, high-divergence genes, namely, genes that respond differently to the microbiomes from the four primate host species, tend to be enriched for categories related to disease, inflammation, and cancer (Figures 3C, S3A, and S3B). Of note, colorectal cancer, rheumatoid arthritis, and *Salmonella* infection functional categories are enriched among high-divergence genes and have all been associated with gut microbiome composition in previous studies (Dahmus et al., 2018; Scher and Abramson, 2011; Ferreira et al., 2011). Moreover, when considering host genes that have been previously associated with complex human traits through genome-wide association studies (GWAS) using data in the GWAS catalog (Buniello et al., 2019), we find that high-divergence genes are enriched with traits and diseases that have also been linked to the microbiome, such as Crohn's disease (CD), inflammatory bowel disease (IBD), and body mass index (Figure 3D; STAR Methods). This might indicate that these complex disease phenotypes may be modulated by differences in composition of the gut microbial community through the regulation of these key host genes.

Next, we sought to identify genes whose response is directly correlated with the abundance of specific microbial taxa. To do so, we used mixed linear models that integrated host response transcriptomic data (via RNA-seq) and microbial species abundance information data (via shotgun metagenomics; see STAR Methods). We identified 25 microbial species that drive the expression of 80 differentially expressed host genes across the four hominids (Figure 4A; Table S7, 162 host

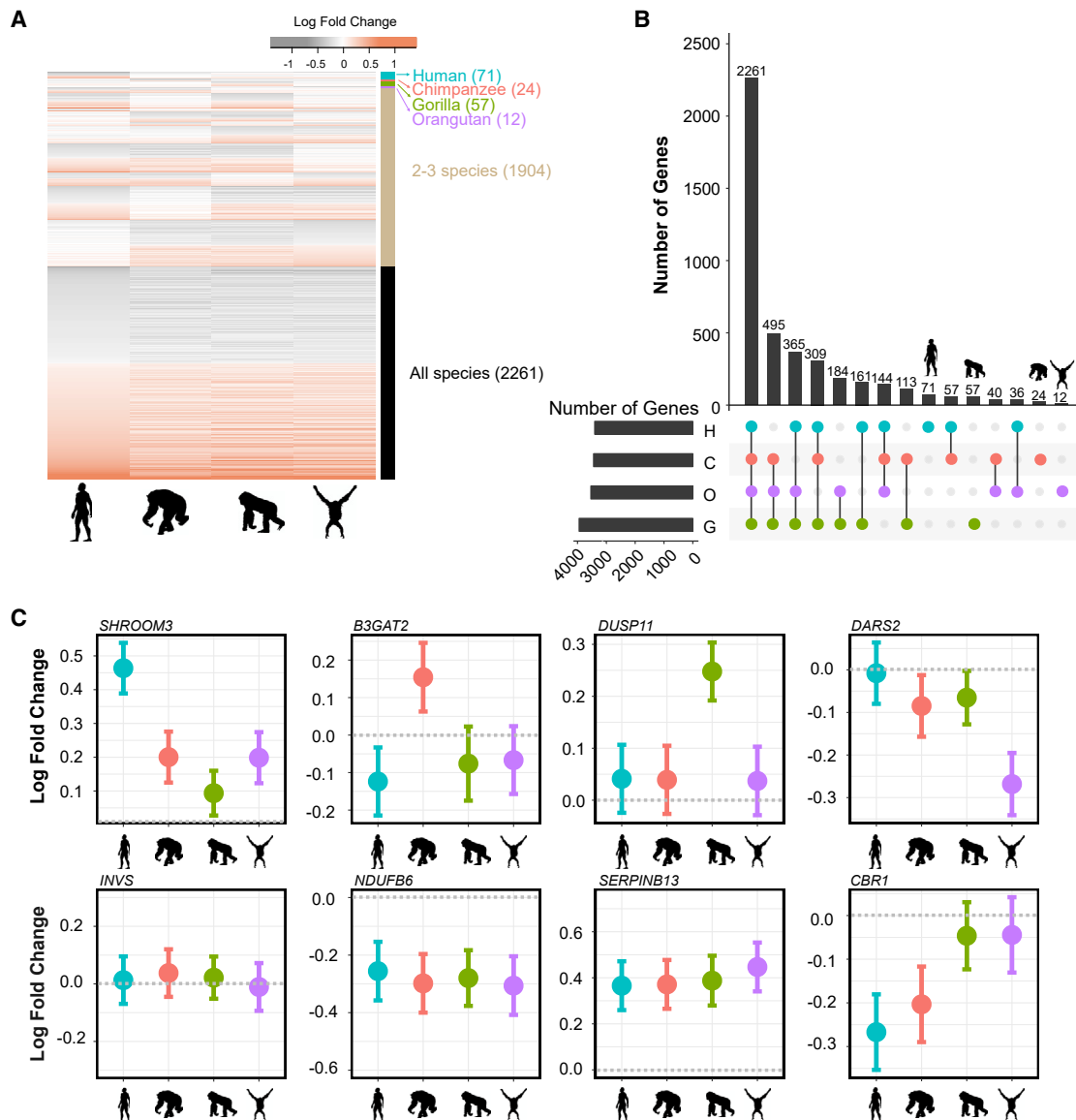


Figure 2. Patterns of host gene expression change in response to hominid microbiome treatment

(A) Heatmap showing the \log_2 fold change for all differentially expressed host genes (rows), grouped by expression pattern. Each row in the heatmap represents a host gene and each column represents one of the four hominid species. The colored bar on the right hand side indicates the hominid microbiome to which these genes respond (Benjamini-Hochberg FDR < 0.1). See also Table S13.

(B) UpSet plot visualizing the intersections among the sets of host genes that respond to hominid microbiomes. The x axis indicates the hominid species (or combination of hominid species) considered in the intersection and is represented by colored dots underneath the x axis (each color depicts a hominid species as in Figure 1). The y axis indicates the number of genes in the intersection. The bars in the lower left indicate the total number of differentially expressed genes by hominid species. For example, the leftmost bar shows that 2,261 genes are differentially expressed in response to microbiomes from all four hominid species, and the rightmost bar shows that 12 genes are differentially expressed in response to only orangutan microbiomes. See also Table S3.

(C) Examples of expression patterns of eight differentially expressed genes. Each panel represents a single host gene, labeled at the top of the plot. The x axis represents the four hominid species, and the y axis represents the \log_2 fold change in expression of the gene. Each dot represents the \log_2 fold change in response to the microbiomes of each hominid, with error bars indicating the SE. The gray dotted line is at zero and denotes no differential expression. See also Tables S3 and S13.

gene-microbial taxon pairs in total, Benjamini-Hochberg FDR < 0.05). A heatmap of the interactions reveals two roughly defined major clusters, one of which includes a subcluster of host genes that are downregulated by microbial taxa that are rare or absent in humans but present in the other hominids, such as *Prevotella*

copri, *Methanobrevibacter* (unclassified), and *Phascolarctobacterium succinatutens* (highlighted in Figure 4A; also see Figure 1D for *P. copri* and *P. succinatutens* abundances across hominids). Genes that are downregulated in the presence of these microbial species are significantly enriched for several immune-related

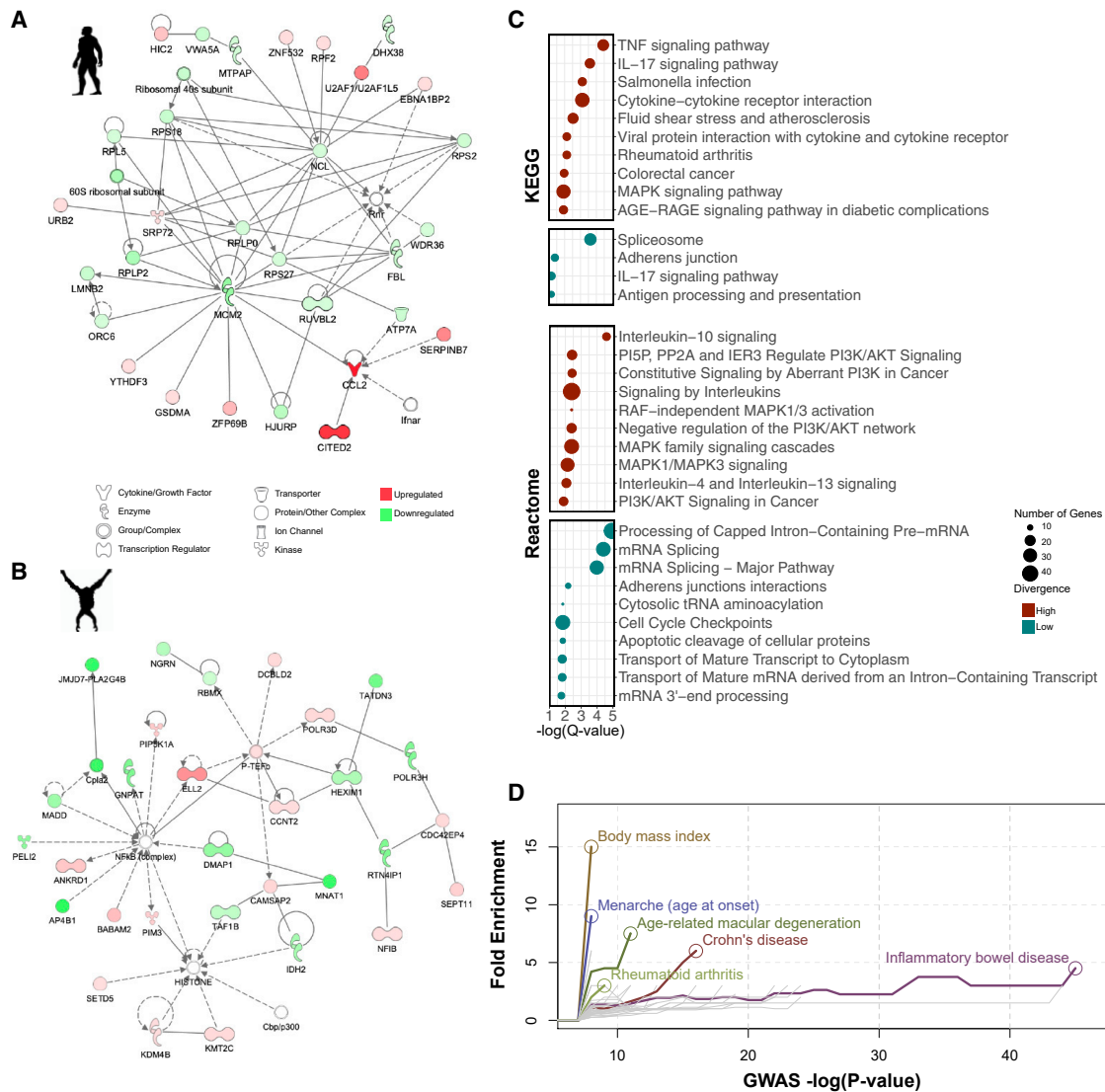


Figure 3. Interaction networks and functional enrichment categories for host genes responding to hominid gut microbiomes

(A) Interaction network showing host genes that respond only to human microbiomes, generated using Ingenuity Pathway Analysis. Each node indicates a gene, and the color of the node indicates whether the gene is upregulated (red) or downregulated (green). The shape of each node represents a specific function as depicted in the legend. A line indicates a direct interaction, and a dashed line indicates an indirect interaction. See also Figure S2.

(B) Similar to (A), but with host genes that respond only to orangutan microbiomes.

(C) Functional categories in the KEGG (top) and Reactome (bottom) databases enriched among high-divergence genes (red) and low-divergence genes (blue). The x axis indicates the statistical significance of enrichment, and the circle size corresponds to the number of genes in each category, as shown in the legend (Benjamini-Hochberg FDR < 0.1). See also Figure S3 and Table S6.

(D) Complex disease enriched among genes that respond to hominid microbiomes. Fold enrichment (y axis) is shown for a given p value threshold (x axis) to define genes that are associated with each complex disease in the GWAS catalog. Each colored line represents a complex disease with a statistically significant enrichment after multiple test correction (using Fisher's exact test, FDR < 0.1; see STAR Methods), with a circle indicating the most significant p value threshold. Diseases that did not reach significance are shown in gray lines.

pathways, such as cytokine activity, interleukin (IL)-7 signaling, malaria, Legionellosis, and tumor necrosis factor (TNF) signaling (Table S8). Using a similar method, we identified 89 microbial pathways that drive the expression of 310 unique host genes for a total of 2,061 significant microbial pathway-host gene interactions (Benjamini-Hochberg FDR < 0.05). For simplicity, we focused on the top 48 microbial pathways that drive the expres-

sion of the top 44 unique host genes (Figure 4B; Table S9), with a total of 216 microbial pathway-host gene pairs (examples of specific interactions can be found in Figure S4A). Clustering of this interaction data revealed three main clusters (I, II, and III), with genes in cluster II associated with pathways that are more abundant in humans compared to other hominid microbiomes. These host genes are enriched in functional categories related to

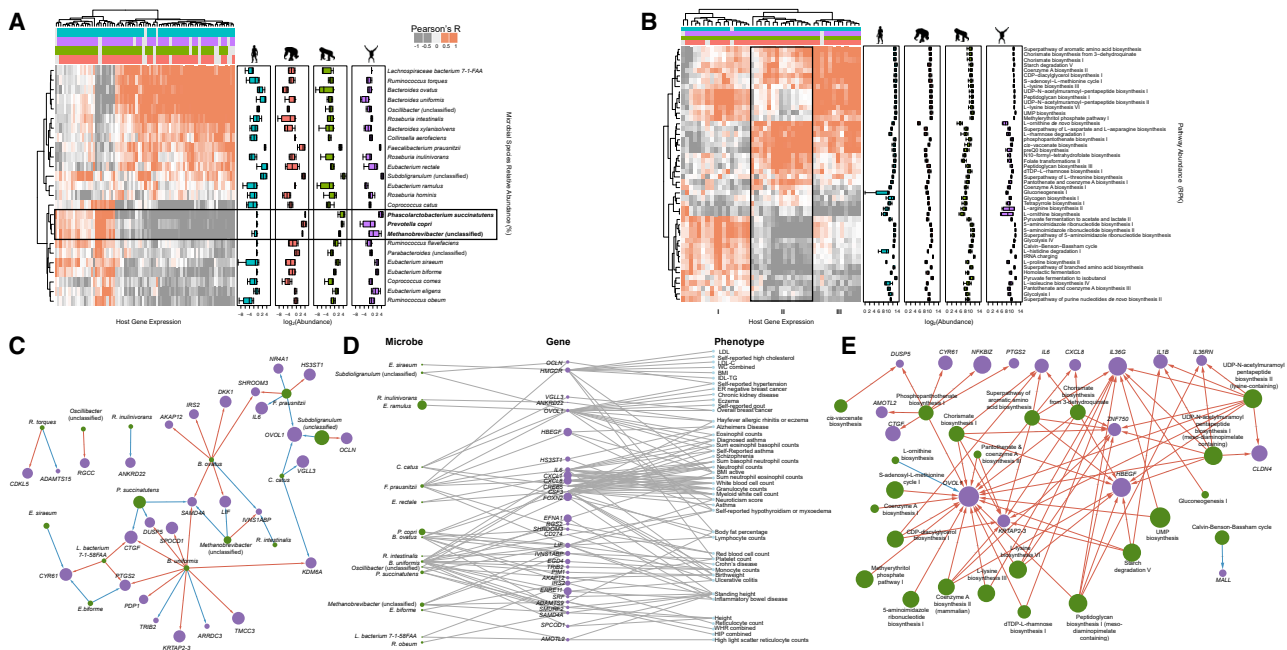


Figure 4. Relationship between host gene expression and specific microbiome features

(A) Heatmap showing correlations between abundance of microbial species from metagenomics sequencing data (rows) and host gene expression response (columns) (Benjamini-Hochberg FDR < 0.05). The colored bars at the top indicate to which hominid microbiome (or combination of hominid microbiomes) a gene responds, with blue, orange, green, and purple bars depicting the human, chimpanzee, gorilla, and orangutan microbiomes, respectively. Boxplots to the right show the abundance of each microbial species in each hominid microbiome (microbial abundance transformed by log₂) using the same color scheme. See also [Figure S4](#) and [Table S7](#).

(B) Similar to (A), but showing the abundance of microbial pathways instead of microbial species (Benjamini-Hochberg FDR < 0.05). See also [Figure S4](#) and [Table S9](#).

(C) Network visualization of high-divergence host genes (purple nodes) and microbial species (green nodes) to which these host genes respond (shown as arrows). The node size of microbial species corresponds to abundance, and the node size of host genes corresponds to log₂ fold change of the differential expression in response to microbiome exposure. Arrow colors indicate whether a microbial species increases (blue) or decreases (red) the expression of the connected host gene. See also [Table S8](#).

(D) Three-tier network showing microbial species (left column), the host genes they each regulate (middle column), and the TWAS phenotypes these genes are associated with (right column). Microbial species and host gene node size indicate microbial abundance and differential expression, respectively, correlated with high-divergence genes and TWAS phenotypes. See also [Table S11](#).

(E) Similar to (C), but showing microbial pathways instead of microbial species. See also [Table S10](#).

inflammation and infectious disease, including Legionellosis, malaria, and pertussis, and overlap with genes found in the cluster described above in the species-level analysis ([Table S10](#)).

To investigate specific host gene-microbe interactions, we considered the network of all high-divergence host genes for which expression is driven by microbial species (28 host genes and 14 microbial species; Benjamini-Hochberg FDR < 0.01). We find that certain microbial taxa are represented in highly connected nodes and likely control the regulation of several high-divergence host genes ([Figure 4C](#)). For example, two *Bacteroides* species, *B. ovatus* and *B. uniformis*, drive the expression of several host genes, including *LIF* and *DUSP5* respectively, both of which have been previously associated with inflammation ([Yue et al., 2015](#); [Habibian et al., 2017](#)). *Bacteroides* is a highly abundant microbial genus in the human gut and is known to have mixed effects on human health ([Wexler, 2007](#)). Notably, *B. ovatus* is highly abundant in the human microbiome samples, but is at low abundances in the orangutan gut microbiomes and entirely absent in the chimpanzee and gorilla microbiomes ([Figure 1D](#), FDR < 0.1).

To explore the possible phenotypic consequences of host genes for which expression is driven by certain microbial species, we integrated gene-trait associations identified through transcription-wide association study (TWAS). TWAS identifies associations between gene expression and complex traits by considering genetically predicted gene expression from expression quantitative trait locus (eQTL) studies and SNP-trait associations from GWAS. We considered genes implicated in 114 complex traits through Probabilistic TWAS ([Zhang et al., 2020](#)) and found that expression of 44 out of 57 high-divergence host genes is associated with 43 complex phenotypes ([Figure 4D](#)). These include diseases and phenotypes previously linked to the gut microbiome, including CD, IBD, ulcerative colitis, body mass index, body fat percentage, and schizophrenia ([Figure 4D](#); [Table S11](#)). We found several microbial taxa that have higher abundance in the non-human microbiomes, including *P. copri* and *P. succinatutens*, which have previously been hypothesized to have protective effects, downregulate the expression of host gene *LIF*, which has been linked to

ulcerative colitis, IBD, and CD in our TWAS analysis (Figures 4C and 4D) (De Vadder et al., 2016; Morgan et al., 2012). These results are consistent with findings from the enrichment analysis reported in Figures 3C and 3D, where we found phenotypes related to inflammation were driven by high-divergence genes. Furthermore, we find that *Eubacterium rectale* and *B. ovatus*, microbes that have higher abundance in humans and that have been previously associated with IBD (Zhang et al., 2017; Noor et al., 2010), upregulate the expression of *CSF3*, which has been reported as upregulated in ulcerative colitis patients (Hotte et al., 2012; de Lange and Barrett, 2015)

To investigate specific host gene-pathway interactions, we constructed a network of the most significant interactions between microbial pathways and high-divergence host genes as described above (Figure 4E; see STAR Methods). We find that nine of these 17 host genes, including *DUSP5*, *CYR61*, *NFKBIZ*, *PTGS2*, *IL6*, *CXCL8*, *IL36G*, *IL1B*, and *IL36RN* (all displayed at the top layer in Figure 4E) have been implicated in immune function or inflammation (Habibian et al., 2017; Emre and Imhof, 2014; Hörber et al., 2016; Müller et al., 2018; Cox et al., 2004; Rincon, 2012; Gales et al., 2013; Wang et al., 2017; Ren and Torres, 2009; Onoufriadis et al., 2011). We found that these genes are associated with several microbial pathways, including phosphopanthothenate biosynthesis I, chorismate biosynthesis, UDP-N-acetylmuramoyl pentapeptide biosynthesis II (lysine-containing), and UDP-N-acetylmuramoyl pentapeptide biosynthesis I (meso-diaminopimelate containing).

DISCUSSION

Interactions between hominid hosts and their microbiomes have been an underexplored area of research, and the complexity of the host-microbiome relationship makes identifying the specific microbial features that causally impact the host phenotype inherently challenging. Here, we use an *in vitro* model to assess how gut microbiomes from different host species impact gene regulation, which is a likely mechanism for microbes to drive changes in host phenotype and health. Inoculating host colonic epithelial cells with live gut microbiome communities from four great ape species, we find that most host genes are regulated similarly by microbiomes from all four hominid microbiomes. However, some host genes are regulated only by microbiomes from a single hominid; these genes are enriched with immunity functions and are involved in the development of IBD.

Chimpanzees, gorillas, and orangutans are our closest extant relatives, making these species an important study system for understanding human evolution as well as the genetic and environmental etiology of human-specific diseases. Distinct physiological, cognitive, and behavioral differences between primate species are hypothesized to be the result of changes in host gene regulation (Gilad et al., 2006; King and Wilson, 1975; Britten and Davidson, 1971; Enard et al., 2002). Indeed, studies have identified genes showing a species-specific expression pattern and genes for which regulation likely evolves under natural selection (Blekhman et al., 2008; Brawand et al., 2011). Here, we show that microbiomes of different hominid species elicit different gene expression responses in the same type of intestinal epithelial cells (human colonocytes). Although we show that most host

genes respond to microbiomes from different hominids in a similar manner, we also identified genes that exhibit a species-specific response. Thus, it may be tempting to hypothesize that some of the species-specific differences in gene expression observed previously are driven by interactions with the gut microbiome. These species-specific microbiome-regulated host genes might facilitate host-specific adaptations to physiological or dietary constraints; for example, our analysis indicates that genes with a response to only orangutan microbiomes are enriched for carbohydrate metabolism, lipid metabolism, and small molecule biochemistry, which suggests that the interaction of the orangutan microbiome and colonic epithelial cells may aid in digestion of specific macronutrients, especially those associated with diets rich in high-energy, highly digestible plant sources (e.g., ripe fruit).

In addition to environmental adaptations, species-specific responses to the microbiota may indicate tightly controlled symbiotic relationships that may result in disease phenotypes when altered. We find that high-divergence genes—namely, genes that respond discordantly to microbiomes from different hominid species—are enriched for traits associated with disease, such as inflammation and aberrant apoptosis. This suggests that genes with a response highly sensitive to the variation across hominid microbiomes may possibly play a role in host disease traits. These genes are also significantly associated with relevant disease traits in the GWAS catalog and in our TWAS analysis, including CD and IBD. Significant distinctions exist in gut microbiome composition and diversity across apes with marked differences in subsistence strategies: for instance, industrialized human societies and primates in captivity have lower gut microbiome diversity and show higher incidences of noncommunicable diseases than small-scale human populations and wild non-human primates, respectively (Gomez et al., 2016a; Clayton et al., 2016). Thus, one hypothesis is that these unique features of the microbiome are causal for the development of diseases common in humans living in industrialized areas, but not in non-industrialized human populations or in non-human wild primates, such as IBD. Our results are consistent with this hypothesis, and further suggest that a mechanism by which the microbiome can affect disease risk is through regulating the expression of host genes in interacting colonic epithelial cells. For example, we found that several microbes that have lower abundance in humans compared to the other hominids, including *P. copri* and *P. succinatutens*, downregulate the expression of the gene *LIF*, which has been associated with IBD (Figure S4B). This suggests that these microbes may confer a protective effect through regulation of host genes, and their absence in humans is possibly detrimental. Conversely, we found that microbes that have higher abundance in humans compared to the other hominids, including *B. ovatus* and *E. rectale*, upregulate the expression of *CSF3*, which has been associated with IBD (Figure S4B). This suggests that these microbes may have a human-specific pathogenic effect. Moreover, some of the genes we found to be regulated by the microbiome in a species-specific manner, such as *IL1B*, *IL6*, *IL36G*, *IL36RN*, and *CXCL8*, have been previously implicated in IBD (Schulze et al., 2008; Khor et al., 2011; Parisinos et al., 2018; Gijsbers et al., 2004; Russell et al., 2016;

Müller et al., 2018), whereas others, such as *DUSP5*, *CYR61*, *NFKB1Z*, and *PTGS2*, have roles in immune response (Habibian et al., 2017; Emre and Imhof, 2014; Hörber et al., 2016; Cox et al., 2004).

In conclusion, we find that gut microbial communities from different hominids mostly elicit a conserved regulatory response in host cells, whereby most host genes respond similarly to hominid microbiomes. However, we also find that some host genes show a divergent response, and a number of host genes respond only to microbiomes from one hominid species and not the others. These genes are enriched in functional categories related to immunity and inflammation and are over-represented in pathways involved in autoinflammatory diseases, such as IBD and CD. These results represent an important step toward understanding the causal relationships between variation in the gut microbiome across hominids and the regulation of intestinal epithelial cells. We hope that future studies will expand on this work using organoid culture or animal models to understand microbial regulation of host genes in more complex systems. Moreover, we expect that testing specific isolates or manipulating the composition of the microbiome will allow characterizing the contribution of specific microbes to the development of disease through regulation of host genes.

Limitations of study

Our ability to interpret these results in a comparative evolutionary context is limited by the unavailability of colonocytes from the non-human hominids in the study. In addition, the non-human hominids in the study are all captive, and our comparison of these samples to publicly available data indicates that the captive chimpanzee and gorilla microbiomes in our study differ from their wild counterparts; nevertheless, the microbiomes used in this study still cluster by host species identity, and preserve clear between-species variation in microbiome composition. Another limitation of our analysis is that the taxonomic profiling of metagenomic shotgun sequencing data relies on databases that are biased toward microbes residing in human microbiomes and might impact our ability to detect and accurately quantify certain microbes in the non-human samples. Moreover, the *in vitro* approach used here represents a simplified version of the complex interactions occurring at the organismal level. Nevertheless, our approach allows for tightly controlled experimental conditions that can be tailored to the specific question of interest, by focusing on the relevant host cell type and microbiomes, and massively reducing confounding effects of cellular composition and the environment. Indeed, our approach allows controlling for various factors that may affect both the microbiome and host gene regulation, such as organismal-level variables (e.g., infection and hormones), host genetic variation, environmental factors (e.g., host diet), and oscillations and circadian dynamics in the microbiome and host gene expression.

STAR★METHODS

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SUPPLEMENTAL INFORMATION

Supplemental information can be found online at <https://doi.org/10.1016/j.celrep.2021.110057>.

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AUTHOR CONTRIBUTIONS

R.B., F.L., and A.L.R. conceived and designed the experiment. A.L.R., A.A., and C.C. performed the experiment. A.G., J.B.C., M.B.B., and K.P. contributed samples and data. A.L.M. analyzed the data, with help from R.P.-R., A.L.R., X.W., and J.R. A.L.M., R.B., and F.L. wrote the manuscript. All the authors read and edited the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Biological samples		
Fecal samples human	OpenBiome	Cat# FMP250, lot# 0028-004-C lot# 0065-0016-D lot# 0110-0006-01 lot# 0111-0014-01 lot# 0112-0002-02
Fecal samples non-human primate	This paper	N/A
Chemicals, peptides, and recombinant proteins		
Poly-l-lysine (PLL)	ScienCell Research Laboratories	ScienCell 0413
Critical commercial assays		
Dynabeads mRNA Direct Kit	Ambion	Cat# 61011
NexteraXT DNA Sample Preparation Kit	Illumina	Cat# FC-130-1005
Deposited data		
Raw data (RNA-seq, metagenomic shotgun, 16S rRNA)	This paper	SRA: PRJNA661048
Novel code	This paper	https://github.com/blekmanlab/Primates https://doi.org/10.5281/zenodo.5589876
Human Reference Genome NCBI Build 38, GRCh38	Genome Reference Consortium	https://www.ncbi.nlm.nih.gov/assembly/GCF_000001405.26/
Experimental models: cell lines		
Human Colonic Epithelial Cells: HCoEpiC, lot: 9763	ScienCell Research Laboratories	lot# 9763
Software and algorithms		
R	N/A	https://www.r-project.org/
FastQC	N/A	https://www.bioinformatics.babraham.ac.uk/projects/fastqc/
Trimmomatic	Bolger et al., 2014	http://www.usadellab.org/cms/?page=trimmomatic
HISAT2	Kim et al., 2019	http://daehwankimlab.github.io/hisat2/
HUMann2	Franzosa et al., 2018	https://huttenhower.sph.harvard.edu/humann2/
Metaphlan2	Truong et al., 2015	https://huttenhower.sph.harvard.edu/metaphlan2/#:~:text=The%20Huttenhower%20Lab%20%3E%20MetaPhlan2,from%20metagenomic%20shotgun%20sequencing%20data.
Diamond	Buchfink et al., 2015	https://www.uppmax.uu.se/resources/databases/diamond-protein-alignment-databases/
DADA2	Callahan et al., 2016	https://benjineb.github.io/dada2/tutorial.html
Vegan	N/A	https://cran.r-project.org/web/packages/vegan/vignettes/intro-vegan.pdf
DESeq2	Love et al., 2014	https://bioconductor.org/packages/release/bioc/html/DESeq2.html
Ingenuity Pathway Analysis	N/A	https://digitalinsights.qiagen.com/products/ingenuity-pathway-analysis

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Continued

REAGENT or RESOURCE	SOURCE	IDENTIFIER
ClusterProfiler	Yu et al., 2012	https://bioconductor.org/packages/release/bioc/html/clusterProfiler.html
bowtie2	Langmead and Salzberg, 2012	http://bowtie-bio.sourceforge.net/bowtie2/index.shtml
MinPath	Ye and Doak, 2009	https://omics.informatics.indiana.edu/MinPath/
UniRef90	Suzek et al., 2015	https://www.uniprot.org/help/uniref

RESOURCE AVAILABILITY

Lead contact

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Ran Blekhman (blekhman@umn.edu)

Materials availability

This study did not generate new unique reagents.

Data and code availability

All sequencing data (RNA-seq, metagenomic shotgun, and 16S rRNA) have been deposited at the Sequence Read Archive (SRA) and are publicly available as of the date of publication, under BioProject accession number SRA: PRJNA661048. Accession numbers are listed in the [Key Resources Table](#).

All original code has been deposited on Github (<https://github.com/blekhmanlab/Primates>) and on Zenodo and is publicly available as of the date of publication. DOIs are listed in the [Key Resources Table](#).

Any additional information required to reanalyze the data reported in this paper is available from the lead contact upon request.

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Sample acquisition and live microbiota extraction

See [Table S1](#) for full details about the human and non-human primate fecal samples used in this analysis. Non-human fecal samples from gorillas and orangutans were collected from captive animals immediately after defecation. One orangutan who donated two samples was on a low dose of antibiotics for chronic colitis. Samples were collected as soon as possible (within an hour of defecation) into a 50mL conical tube containing 20mL of cryoprotectant solution consisting of a 50:50 mixture of glycerol and saline solution. The cryoprotectant was filter sterilized through a 0.22 μ m filter. Samples were shaken vigorously to distribute the cryoprotectant. Gorilla and orangutan samples were stored at -80°C within 1 hour after collection and shipped to the lab on dry ice. Chimpanzee samples were stored at -20°C within 1 hour of collection and then shipped to the U.S. lab on dry ice within one day. Human fecal samples were purchased from OpenBiome and arrived frozen on dry ice. The following briefly describes the protocol by which OpenBiome processes stool samples. The sample is collected by OpenBiome and given to a technician within 1 hour of defecation. The mass of the sample is measured and transferred to a sterile biosafety cabinet. The stool sample is put into a sterile filter bag, and a sterile filtered dilutant of 12.5% glycerol is added with a normal saline buffer (0.90% [wt/vol] NaCl in water). The sample solution is then introduced to a homogenizer blender for 60 s and aliquoted into sterile bottles. The bottles are then immediately frozen at -80°C . Any sample not fully processed within 2 hours of passage is destroyed.

To extract fecal microbiota from the non-human primate samples, inside a sterile low-oxygen cabinet we placed fecal material into a sterilized disposable standard commercial blender cup, and added 20mL glycerol to reach approximately 30mL glycerol and 200mL normal saline buffer (0.90% [wt/vol] NaCl in water). Fecal material was blended until fully homogenized (about 1-2 min). Blended material was transferred to the same side of the membrane in a 330-micron filter bag and the liquid suspension of the bacterial community was collected on the other side of the filter. The resulting microbiota suspension was then mixed and aliquoted into small tubes and stored at -80°C .

The research and sample collection in this study complied with protocols approved through the University of Minnesota Institutional Animal Care and Use Committee.

Colonic Epithelial Cell Line and Growth Conditions

Experiments were conducted using primary human colonic epithelial cells (HCoEpiC, lot: 9763), hereby called colonocytes (ScienCell Research Laboratories, Carlsbad, California, USA, 2950). The cells were cultured on plates or flasks coated with poly-L-lysine (PLL),

according to the supplier's specifications (ScienCell 0413). Colonocytes were cultured in colonic epithelial cell medium supplemented with colonic epithelial cell growth supplement and penicillin-streptomycin according to the manufacturer's protocol (ScienCell 2951) at 37°C with 5% CO₂. At 24 hours before treatment, cells were changed to antibiotic-free medium and moved to an incubator at 37°C, 5% CO₂, and a reduced 5% O₂.

METHOD DETAILS

Colonocyte with hominid-derived microbiota treatment experiment

The experimental protocol used for the treatment of colonocytes with microbiota has previously been described in [Richards et al. \(2016\)](#). Fecal microbiota were not thawed until the day of the experiment. Prior to treatment, the microbiota was thawed at 30°C, and the microbial density (OD₆₀₀) was assessed via a spectrophotometer (Bio-Rad SmartSpec 3000). Medium was removed from the colonocytes and fresh antibiotic-free medium was added to the cells, with a final microbial ratio of 10:1 microbe:colonocyte in each well. Additional wells containing only colonocytes were also cultured in the same 24-well plate for use as controls.

After 2 hours, the wells were scraped on ice, pelleted, and washed with cold phosphate-buffered saline (PBS) and then resuspended in lysis buffer (Dynabeads mRNA Direct kit, ThermoFisher Scientific, Waltham, Massachusetts, USA) and stored at –80°C until extraction of colonocyte RNA for RNA-seq. We conducted both metagenomic shotgun sequencing and 16S rRNA sequencing on the microbiomes at four points: before preparation (raw), after preparation (prepared), cultured with colonocytes (colonocytes) and cultured without colonocytes (control). Previous experiments have shown that microbiome composition does not change drastically over the 2 hour co-culture period ([Richards et al., 2019](#)). Human fecal microbiome samples were purchased as “prepared” from Openbiome and therefore were not sequenced raw.

RNA-seq experiment and data processing

Poly-adenylated mRNA was isolated from thawed cell lysates using the Dynabeads mRNA Direct Kit (Ambion) following the manufacturer's instructions. RNA-seq libraries were prepared using a protocol modified from the NEBNext Ultradirectional (NEB) library preparation protocol to use Barcodes from BIOO Scientific added by ligation, as described in [Richards et al. \(2019\)](#). The libraries were then pooled and sequenced on two lanes of the Illumina Next-seq 500 in the Luca/Pique-Regi laboratory using the high output kits for 75 cycles to obtain paired-end reads. Reads were 80 bp in length. Read counts ranged between 12,632,223 and 36,747,968 reads per sample, with a mean of 18,726,038 and median of 16,993,999 reads per sample.

FastQC was used to determine the quality of reads from raw data (FastQC, version 0.11.5). Trimmomatic was used to trim adapters. FastQC was again used to determine the quality of reads after trimming of adapters (Trimmomatic version 0.33). Transcripts were aligned to database GRCh38 using HISAT2 (HISAT2 version 2.0.2) ([Kim et al., 2019](#)). After alignment, read counts ranged between 10,817,737 and 33,592,529 aligned reads per sample, with a mean of 17,142,585.72 and a median of 15,542,693.5 aligned reads per sample. Overall, the average alignment rate was ~70% across samples ([Figure S5](#)). The R ‘Subread’ package with the ‘featureCounts’ program was used to make the transcript abundance file (R version 3.3.3, Subread version 1.4.6).

16S rRNA sequencing

Sequencing on the 16S rRNA V4 region was performed at the University of Minnesota Genomics Center using the protocol described in [Gohl et al. \(2016\)](#). DNA isolated from fecal samples was quantified with qPCR and the V4 region of the 16S rRNA gene was amplified using PCR with barcodes for multiplexing.

The forward indexing primer sequence is - **AATGATACGGCGACCACCGAGATCTACAC**[i5]TCGTCCGCGAGCGTC and the reverse indexing primer sequence is - **CAAGCAGAAGACGGCATACGAGAT**[i7]GTCTCGTGGGCTCGG (where the bolded regions are the p5 and p7 flow cell adapters and [i5] and [i7] refer to the index sequence codes used by Illumina). The qPCR step starts with an initial denaturing step at 95°C for 5 min followed by 35 cycles of denaturation (20 s at 98°C), annealing (15 s at 66°C) and elongation (1 min at 72°C). After qPCR, samples are normalized to 167,000 molecules/μl. This is based on the volume of sample used for PCR1 (3μl), so 500,000 molecules is roughly 10x the target sequencing coverage. The next PCR (PCR1) step is similar to the qPCR step, except with only 25 cycles of denaturation, annealing, and elongation. After the first round of amplification, PCR1 products are diluted 1:100 and 5μl of 1:100 PCR1 is used in the second PCR reaction. The next step (PCR2) is similar to the previous two PCR protocols, except with only 10 cycles of denaturation, annealing, and elongation. Next, pooled samples were denatured with NaOH, diluted to 8 pM in Illumina's HT1 buffer, spiked with 15% PhiX, and heat denatured at 96°C for 2 minutes immediately prior to loading. A MiSeq 600 cycle v3 kit was used to sequence the sample. The following Nextera adaptor sequences for post-run trimming are also used. For read 1 - CTGTCTTATACATCTCCGAGCCCACGAGACNNNNNNNNATCTCGTATGCCGTCTTCTGCTTG and for read 2 - CTGTCTTATACATCTGACGCTGCCGACGANNNNNNNNGTGTAGATCTCGTGGTCCGATCATT

Metagenomic shotgun sequencing

Metagenomic shotgun sequencing on prepared microbiota samples was performed at the University of Minnesota Genomics Center (UMGC). DNA samples were quantified using a fluorimetric PicoGreen assay gDNA samples were converted to Illumina sequencing libraries using Illumina's NexteraXT DNA Sample Preparation Kit (Cat. # FC-130-1005). 1 ng of gDNA was simultaneously fragmented and tagged with a unique adaptor sequence. This “tagmentation” step is mediated by a transposase. The tagmented DNA was

simultaneously indexed and amplified with 12 PCR cycles. Final library size distribution was validated using capillary electrophoresis and quantified using fluorimetry (PicoGreen). Truseq libraries were hybridized to a NextSeq. Base call (.bcl) files for each cycle of sequencing were generated by Illumina Real Time Analysis (RTA) software. The base call files were demultiplexed and then converted to index specific fastq files using the MiSeq Reporter software on-instrument.

QUANTIFICATION AND STATISTICAL ANALYSIS

Characterizing microbiota

To identify microbial features from the metagenomic shotgun sequencing data, including taxa and pathway abundances, we used the HUMAnN2 pipeline with Metaphlan2 (HUMAnN2 v0.11.1, Metaphlan2 v0.2.6.0) (Truong et al., 2015; Franzosa et al., 2018). FastQC v0.11.7 was used to determine the quality of sequencing reads before trimming. Sequencing adapters were trimmed from the raw reads using Trimmomatic (Trimmomatic v0.33) (Bolger et al., 2014). FastQC v0.11.7 was again used to determine quality of sequencing reads after trimming the sequencing adapters from the reads (Figure S6). Metaphlan2 was used to assign taxonomy at all taxonomic levels to the sequencing reads in each sequencing file, and in particular to get relative abundances of microbial taxa for each sample. The HUMAnN2 pipeline utilizes bowtie v0.2.2 for read alignment (Langmead and Salzberg, 2012), DIAMOND v0.8.22 for high throughput protein alignment (Buchfink et al., 2015), MinPath (Ye and Doak, 2009) for pathway reconstruction from protein family predictions. The UniRef90 database was used for determining gene family abundances (Suzek et al., 2015). We found a total of 166 named microbial species detected in at least one sample (Figure S7).

Principal Coordinate Analysis of Samples

Using the 16S rRNA data from the fecal microbiota samples, we used the R package 'DADA2' (DADA2, version 1.2.2) to identify amplicon sequence variants (ASVs) from the reads (Callahan et al., 2016). DADA2 was used to filter and trim sequences from raw reads. Forward reads were trimmed to position 240 and reverse reads were trimmed to position 160. Reads were truncated at the first quality score less than or equal to 2. Reads with more than two errors were discarded after truncation. Amplicon sequences were dereplicated using the function 'derepFastq.' Sample composition was inferred using the 'dada' function. Chimeras were removed using 'removeBimeraDenovo.' We assigned taxonomy to the resulting ASVs using 'assignTaxonomy.' Using the R package 'vegan' (version 2.5-3), we calculated Bray-Curtis dissimilarities and plotted these as a principal coordinate analysis plot (Figure 1B).

Comparison of microbiome composition between captive and wild gorillas and chimpanzees

Previous studies have shown differences in composition between wild and captive primate microbiomes (Clayton et al., 2016). To compare our captive primate samples to their wild counterparts, we downloaded the processed ASV table from Campbell et al. (2020), which sequenced gut microbiomes from wild chimpanzees and wild gorillas. Campbell et al. (2020) sequenced wild gorilla and chimpanzee fecal microbiome samples using the Illumina MiSeq, targeting the V4 region of the 16S rRNA gene. Campbell et al. (2020) processed their data using DADA2, v1.8, and assigned taxonomy on the resulting ASVs using the Greengenes database. The data in our study was sequenced on the same region and processed with DADA2, v1.2.2, and also assigned taxonomy using Greengenes. Using the R package 'phyloseq', we combined the two datasets and created a stacked barplot showing the taxonomic relative abundances at the phylum level for each of the samples, with captive and wild individuals from each species (Figure S1B).

Species-specific differential expression analysis

We filtered the RNA-seq counts table so that we only consider protein coding genes, reducing the number of considered genes from 60,674 to 19,715. Host genes were filtered for only protein coding genes using the R package 'biomaRt' with ensembl build 37. Within DESeq2 (DESeq2 version 1.14.1), RNA-seq counts were further filtered such that each gene had to be present at least once over all the samples, leaving 17,860 tested genes (Love et al., 2014). DESeq2 uses a negative binomial model to model the count data while it also estimates an appropriate size factor to normalize each sample by its sequencing depth. Additionally, the overdispersion parameter governing the negative binomial distribution is estimated per each gene and using a regularization approach that can monitor outliers and adjust for the mean-variance dependency. The parameter governing the mean gene expression after adjusting to its sequencing depth is modeled as a linear combination that incorporates known batch effects (i.e., plate) and the effect of the biological variable of interest (i.e., each microbiome):

$$\text{Host gene expression} \sim \text{ExperimentPlate} + \text{Microbiome effects.}$$

or, in mathematical terms:

$$Y_{nj} = \sum_s \beta_{js}^M M_{ns} + \sum_p \beta_{jp}^P P_{np}$$

Where Y_{nj} represents the internal DEseq parameter for mean gene expression for gene j and experiment n , M_{ns} is the treatment indicator (control or microbiome for species s), and the β_{js}^M parameter is the microbiome effect for each species. To model plate as a known batch effect we use P_{np} and β_{jp}^P for the plate indicator variable and its effect on gene expression.

For four hominid microbiomes, $2^4 = 16$ effect configurations are possible (for each species combination of which parameters $\beta_{js}^M = 0$), and we ran a likelihood test for each configuration L_j : a gene can respond to a single primate microbiome (chimp, gorilla, human, or orangutan), a gene can respond to two of the four primate microbiomes (chimp-gorilla, chimp-human, chimp-orangutan, gorilla-human, gorilla-orangutan, human-orangutan), a gene can respond to three of the four primate microbiomes (chimp-gorilla-human, chimp-gorilla-orangutan, chimp-gorilla-orangutan, or gorilla-human-orangutan), a gene can respond to all four primate microbiomes, or a gene can show no response to any of the four primate microbiomes. The no-response case is considered the base case, or null model for all the likelihood ratio tests performed.

To identify genes that respond to microbiomes from a specific primate species and to detect the total number of differentially expressed genes that respond to each of the fifteen possible non null combinations of primate microbiomes we ran a likelihood ratio test against the base model, which assumes that the host gene shows no response:

$$\text{Host gene expression} \sim \text{Experiment Plate}$$

and all the coefficients are zero. After determining across all genes and configurations which were statistically significant at $\text{FDR} < 10\%$. We used the likelihood statistics L_{ji} for each gene j and configuration i to calculate the most probable configuration $P(H_{ji}|D) = \frac{L_{ji}}{\sum_i L_{ji}}$.

Simplified models for pairwise comparisons of primate microbiomes

We performed an additional analysis to determine which genes are differentially expressed between each of the pairwise combinations of hominids. We ran the following DESeq2 model:

$$\text{Gene Expression} \sim \text{Experiment Plate} + \text{Species}$$

Where Species is a vector indicating which hominid species the microbiome sample originated from. We filtered the RNA-seq counts so that only genes that are present in at least 3 individuals with count > 0 are considered in the model. We ran contrasts in DESeq2 for all six of the pairwise comparisons between hominids: Chimpanzee-Gorilla, Chimpanzee-Human, Chimpanzee-Orangutan, Gorilla-Human, Gorilla-Orangutan, and Human-Orangutan (Figure S8; Table S12).

Enrichment analysis

Enrichment analysis was performed using Ingenuity Pathway Analysis (IPA, QIAGEN Inc., <https://digitalinsights.qiagen.com/products/ingenuity-pathway-analysis>). We analyzed genes that show a response to microbiomes from a specific primate species. Here, we define those genes as genes that are upregulated or downregulated in response to a specific primate host species, or that show no response to microbiomes from that primate species and show a response to the other three primate host species. For example, genes that show a response only to human microbiomes will be upregulated or downregulated in response to human microbiomes, or show no response to human microbiomes and a response to chimpanzee, gorilla, and orangutan microbiomes. Genes that show a response to three species but not the fourth are also showing a species specific response to the fourth primate species.

We further validated these results using the R package 'ClusterProfiler' for enrichment analysis using all detected genes present in at least one sample as the background set (ClusterProfiler v3.2.14) (Figure 3C) (Yu et al., 2012). We used ENRICH for enrichment analysis of the high and low-divergence genes and extracted the top ten response categories from the GO Biological, GO Molecular, KEGG, and Reactome databases) (Figure S3) (Kuleshov et al., 2016; Chen et al., 2013).

To identify enrichment of high-divergence genes among genes that were previously found to be associated with complex human disease and traits, we used data from the GWAS catalog (Buniello et al., 2019). Since each GWAS has a different distribution of p values and significance cutoffs, we chose to use a set of $-\log_{10}(p \text{ value})$ cutoffs in the range of 8-50 (plotted along the x axis in Figure 3D). For a given trait, we identified the overlap between the genes significantly associated with the disease at each cutoff and high-divergence genes, and calculated a fold enrichment (plotted along the y axis in Figure 3D), defined as the ratio of observed/expected overlap between the two gene sets. We used a Fisher's Exact Test to calculate a p value for each cutoff, and traits for which this value was significant after multiple test correction ($\text{FDR} = 0.1$) were marked with a colored line in Figure 3D.

K-means clustering was performed using the 'kmeans' function in base R (version 3.3.3) on the cluster of microbes *P. copri*, *Methanobrevibacter* and *P. succinatutens* for the genes in Figure 4A. Enrichment analysis was performed using ENRICH on the two clusters of genes. A k-means clustering analysis was also performed on the full set of microbial pathway-host gene correlations in Figure 4B to produce three clusters of genes.

Log fold change of genes by primate species

To calculate the fold changes for each gene for each of the four primate species, we used a similar DESeq2 model to the one described above:

$$\text{Gene expression} \sim \text{ExperimentPlate} + \text{Species}$$

or, in mathematical terms:

$$Y_{nj} = \sum_s \beta_{js}^S M_{ns} + \sum_p \beta_{jp}^P P_{np}$$

Here, Species is a vector indicating which primate species the microbiome sample originated from, and ExperimentPlate controls for the batch effect as before, but we just test the marginal effect of each species-specific parameter β_{js}^S being not different than the untreated control β_j^C . We use the contrast argument in DESeq2 to extract comparisons of each primate species against the control. Thus, this resulted in log fold change calculations for each gene as it responds to each of the four primate species' microbiomes. These values are available in [Table S13](#).

Divergence scores for differentially expressed, conserved genes

Using DESeq2, we identified genes that responded to microbiome treatment. We used the following model to determine whether a gene responds to treatment:

$$\text{Gene expression} \sim \text{ExperimentPlate} + \text{Treatment}$$

Where ExperimentPlate controls for the batch effect of the experiment, and Treatment is a binary vector indicating whether the colonocytes are treated with a microbiome or act as a control for the experiment. Mathematically:

$$Y_{nj} = \mu + \beta_j^T T_n + \sum_p \beta_{jp}^P P_{np}$$

Where Y_{nj} represents the internal DESeq mean gene expression parameter for gene j and experiment n as before, T_n is the treatment indicator (control = 0 or microbiome = 1), and the β_j^T parameter is the microbiome effect. Plate effects are modeled as before. To model plate as a known batch effect we used P_{np} and β_{jp}^P for the plate indicator variable and its effect on gene expression.

Log fold changes for each gene were calculated as described above, and then used to calculate a divergence metric for each gene. We used a similar divergence calculation as described in [Hagai et al. \(2018\)](#). Namely, for the genes identified as responding to treatment with microbiomes, we used the log fold changes for each species in the following equation:

$$\text{Divergence} = \log_2 \left[\frac{1}{6} \sum_i (\log FC \text{ primate}_i - \log FC \text{ primate}_j)^2 \right]$$

Following [Hagai et al. \(2018\)](#), the top 25% of genes were assigned a "high-divergence" status, and the lowest 25% of genes were assigned a "low-divergence" status. These genes were used in the enrichment analyses described below.

The rest of the genes are considered "medium divergence" genes. These genes are used in the enrichment analysis as a background set ([Figures 3A and 3C](#)).

Pairwise correlations between host genes and microbial species and pathways

Using the microbial species abundances calculated from the metagenomic shotgun sequencing, we ran correlation analysis between genes that are differentially expressed with respect to treatment with microbiota and abundances of microbial species. Within DESeq2 (DESeq2 version 1.14.1), RNA-seq counts were further filtered such that each gene had to be present at least twenty times leaving 15,855 tested genes ([Love et al., 2014](#)). Metaphlan2 reports microbial species as a proportion of the total microbial community per sample. Microbial species were filtered such that only microbial species present in at least half of the samples and that reached a total summed relative abundance of 9% were included in the analysis, leaving 36 microbial species. We applied a center log-ratio transformation to the filtered microbial species abundance data. Microbial pathways were filtered such that the total of each pathway had to be greater than a summed threshold of 8000 reads per kilobase (RPK), leaving 95 microbial pathways to be included in the analysis. Microbial pathways were normalized using the centered log ratio transformation in a similar manner to the microbial species.

Using DESeq2, we identified which microbial species or pathways are associated with differentially expressed genes using the following model:

$$\text{Gene Expression} \sim \text{ExperimentPlate} + \text{Treatment} + \text{Microbial feature abundance}$$

Mathematically:

$$Y_{nj} = \mu + \sum_p \beta_{jp}^P P_{np} + \beta_j^T T_n + \beta_j^{A(f)} A_{fn}$$

Where Y_{nj} represents the internal DESeq parameter for gene expression for gene j and experiment n as before, T_n is the treatment indicator (control = 0 or microbiome = 1), and the β_j^T parameter is the microbiome effect. Plate effects are modeled as before. The parameter $\beta_j^{A(f)} A_{fn}$ is used to model the effect of the microbiome feature (i.e., microbial species or pathway) f on gene expression. We statistically test effect $\beta_j^{A(f)} \neq 0$ in a separate DESeq model run for each feature f . We used an FDR correction on the combined

results from all models. The microbial species abundance is a continuous numeric value that represents that center log ratio transformed relative abundance of the microbial feature $f A_m$ for each sample n .

TWAS analysis

To directly investigate whether discovered effects on gene expression may contribute to complex traits, we considered PTWAS gene-trait associations for 114 traits from [Zhang et al. \(2020\)](#). PTWAS utilizes probabilistic eQTL annotations derived from multi-variant Bayesian fine-mapping analysis of eQTL data across 49 tissues from GTEx v8 to detect associations between gene expression levels and complex trait risk. Using the host genes that were highly correlated with a microbial species and fell into the high-divergence category (FDR < 0.05), we overlapped the significant results with genes causally implicated in complex traits across all tissues by [Zhang et al. \(2020\)](#) (PTWAS scan, 5% FDR). We repeated the same analysis with the host genes that were highly correlated with a microbial pathway (FDR < 0.01) and fell into the high-divergence category.