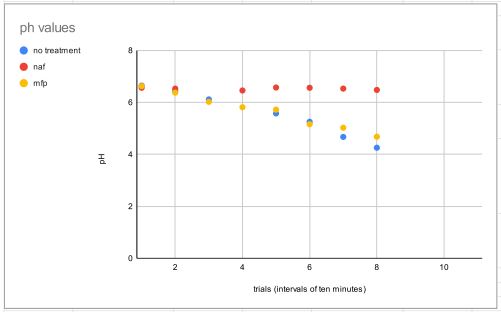
EXPERIMENTAL DATA:

ph drop												
trials	no treatment	naf	mfp	new hand held	1.038575668	6.81689911		6.74:7.00	0.96285714		7/6.74	1.038575668
1	6.646	6.5637	6.62611276		6.480712166						6.646884273	
2	6.439169139	6.53264095	6.376854599				new proportion to multiply all by is					
3	6.117210682	6.82 (void)	6.023738872	switched ph meter here, for naf and further down				1.96285714				
4	6.14 (void)	6.459940653	5.816023739	5.6								
5	5.577151335	6.574183976	5.722551929									
6	5.255192878	6.56379822	5.161721068									
7	4.673590504	6.53264095	5.026706231									
8	4.258160237	6.480712166	4.68		concerned if the bacteria are stocking up							
9						media	PH		optical density = 0.538			
10					n	media before wa	3.84		adding 0.2 of sucrose			
11						AS	6.74					

GRAPH:



NOTES:

DURING	AFTER	BEFORE
after first wash, pellets formed very nicely no film around them, very stuck together and to side of tube which made it easy to decant		
bacteria upon arrival in the media was very prominent and abundant while mixed. once taken out of incubator the bacteria began to clump quickly in a sucrose like fashion even after resuspension		
ph electrode does not go haywire if you rinse it then place it in what you are measuring, then press measure		
visible difference in bacterial growth between the three flasks. the most different is none, which is consistent with the fact that it is dropping so fast. the other two treatments are experiencing a drop, but more incremental, rather than the way no treatment has been showing. need to fix the measurement of bacteria		
no treatment and mfp might still be growing, which might make the ph drop more. we want to be using sub lethal amounts of naf. if our experiment works. no treatment should drop and naf should stay the same.		
intermediate was previously used, initial drop off cou		
ph problems are awful		
	FOR THE FUTURE	
	need to figure out better how to quantitatively measure bacteria out, as we think this time maybe the no treatment had slightly more than the other two with treatments. we will use a little beaker after resuspension with a stir rod and take the bacteria that way, so it can continue to be fully emulsed and not clump in between measuring	
	for next week, go back to laptg without sucrose because thought the bacteria are stocking up on sucrose and dont care about the mpf	
	makes lactate inside the cell not lactic acid - glycosis process when break down carbohydrates	
	look at how much flouride and mpf go inside the cell through F19	